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Past, current and potential future distributions of unique genetic diversity in a cold-adapted mountain butterfly

Short running title less than 40 characters: Climate change and insect genetic diversity

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Biosketch

Melissa Minter is a PhD researcher in the Biology Department at the University of York, UK. Her research examines the genetic diversity of retracting northern and montane butterflies under climate change.

Author contributions

M.M., K.K.D., C.D.T., M.D.M., A.T., T.S. and J.K.H. conceived the ideas; M.M., S.S. and T.S. collected the data; M.M. analysed the data; M.M. drafted the manuscript; all authors contributed to the final manuscript.

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Abstract (300 words)

Aim Climatic changes throughout the Pleistocene have strongly modified species distributions. We examine how these range shifts have affected the genetic diversity of a montane butterfly species, and whether the genetic diversity in the extant populations is threatened by future climate change.

Location Europe.

Taxon *Erebia epiphron* Lepidoptera: Nymphalidae.

Methods We analysed mtDNA to map current genetic diversity and differentiation of *E. epiphron* across Europe to identify population refugia and post-glacial range shifts. We used species distribution modelling (SDMs) to hindcast distributions over the last 21,000 years to identify source locations of extant populations, and to project distributions into the future (2070) to predict potential losses in genetic diversity.

Results We found substantial genetic diversity unique to specific regions within Europe (total number of haplotypes = 31, number of unique haplotypes = 27, $H_d = 0.9$). Genetic data and SDM hindcasting suggest long-term separation and survival of discrete populations. Particularly high rates of unique diversity in post-glacially colonised sites in England ($H_d = 0.64$), suggests this population was colonised from a now extinct cryptic refugium. Under future climate change, SDMs predict loss of climate suitability for *E. epiphron*, particularly at lower elevations (< 1000 metres above sea level) equating to 1 to 12 unique haplotypes being at risk under climate scenarios projecting 1 °C and 2-3 °C increases respectively in global temperature by 2070.

Main conclusions Our results suggest that historical range expansion and retraction processes by a cold-adapted mountain species caused diversification between populations, resulting in unique genetic diversity which may be at risk if distributions of cold-adapted species shrink in future. Assisted colonisations of individuals from at-risk populations into climatically-suitable unoccupied habitat might help conserve unique genetic diversity, and translocations into remaining populations might increase their genetic diversity and hence their ability to adapt to future climate change.

Keywords

Refugia, climate change, Last Glacial Maximum, mountain systems, butterfly, genetic diversity

Data availability statement

Genbank accession numbers for each mtDNA COI sequence can be found in supporting information Appendix S1 in Supporting Information. (All sequences will be uploaded to Genbank upon acceptance.)

1. Introduction

Projecting the future geographic distribution of genetic variation within species' ranges, and the potential loss of genetic variation from anthropogenic climate change, requires understanding of the past, present and future distributions of species (Wroblewska & Mirski, 2018). Geographic variation in the distribution of genes across a species' range results from a combination of historical and current conditions, which influence patterns of genetic differentiation among populations that are, or have been, geographically isolated, and from colonisation bottlenecks during range shifts (Hewitt, 2004). These range shifts and their genetic consequences have primarily been driven by the fundamental niche of a species, or their 'climate-envelope', and species' ranges shift to track environmental changes, altering the location of populations and their genetic structure (Hewitt, 2004; McCallum, Guerin, Breed, & Lowe, 2014; Thomas, 2010). The Earth has gone through many climate fluctuations, including glaciations in the Pleistocene and human-induced climate change in the current Anthropocene (Hewitt, 2004; Santer et al., 2019). Future anthropogenic climate warming may further impact species through distribution changes, genetic erosion and extinctions (Botkin et al., 2007). Cold-adapted/mountain species may be especially vulnerable to future climate changes as they are already restricted to mountain ecosystems where suitable climate space is limited, and loss of genetic diversity within these range-restricted cold-adapted species may reduce their ability to adapt to future changes (Elsen & Tingley, 2015). Understanding how past climatic changes have impacted current genetic structure may allow us to make predictions for the likely extent of genetic loss under future climate change, and thereby prioritise at-risk populations for conservation management (McCallum et al., 2014; Wroblewska & Mirski, 2018).

During the last ice age, ice sheets were at their greatest extension 20,000-21,000 years ago, during the last glacial maximum (LGM) (Crowley & North, 1991; Ray & Adams, 2001). During the LGM, species were thought to persist where climatic conditions were buffered, at lower elevations or in more southerly regions (Dapporto et al., 2019; Morelli et al., 2016), however some studies have shown evidence of species surviving in northern isolated refugia (Provan & Bennett, 2008; Schmitt & Varga, 2012; Stewart & Lister, 2001). Cool-adapted species which currently occur in mountain ecosystems were probably more widespread during the LGM and only became isolated in their current interglacial populations after climate-induced range retraction, although some cold-adapted species were already restricted to isolated glacial refugia during the LGM (Schmitt, 2009; Schmitt, Hewitt, & Muller, 2006). The consequences of past distribution changes will be reflected in current genetic diversity, because contractions and expansions from long-term refugia leave a genetic signature of high diversity in refugia compared to lower diversity in recently-colonised populations (Hewitt, 2000; Keppel et al., 2012; Morelli et al., 2016). Thus understanding historical interactions of cold-adapted species with climate can help us understand current genetic structure and diversity of populations.

Lepidoptera are poikilothermic and therefore sensitive to changes in climate, and those species which are cold-adapted are particularly vulnerable to warmer conditions (Deutsch et al., 2008; Elsen & Tingley, 2015). Some cold-adapted Lepidoptera are experiencing extinctions at their low latitude/elevation margins as the climate deteriorates for these species (Franco et al., 2006; Wilson, Gutierrez, Gutierrez, & Monserrat, 2007). The Mountain Ringlet *Erebia epiphron* is a butterfly found

in the mountains of continental Europe and Britain, and its distribution has retracted 130-150 m uphill in Britain over the past five decades due to climate warming (Franco et al., 2006). Therefore *E. epiphron* represents a good model organism to understand how past climate-induced changes have impacted current genetic structures of populations, and whether genetic diversity may be lost with further climate-induced local extinctions.

Species distribution models (SDMs) are commonly used to project future distributions of species under climate change scenarios (Guo et al., 2017; Urban, 2015), and to develop climate adaptation conservation strategies. These modelling approaches have also been used with palaeoclimate data to hindcast past distributions and to understand how they shape current population structures (Smith, Gregory, Anderson, & Thomas, 2013). Phylogeography with genetic techniques can be used to identify divergence between populations and to infer historical distribution patterns and colonization routes (Luquet et al., 2019). Previous studies have shown how a combination of species distribution modelling and phylogeography can provide better understanding of past, present and future distributions of species, and predict the potential loss of genetic diversity resulting from climatic warming (Schmitt, Habel, Rodder, & Louy, 2014; Wroblewska & Mirski, 2018; Yannic et al., 2014).

In this study, we use mtDNA sequencing to map the current distribution of genetic diversity of the cold-adapted butterfly, *E. epiphron*, and also use species distribution modelling to project current, past, and future distributions of the species. We use this genetic and modelling information to determine the distribution of *E. epiphron* in continental Europe during the last glacial maximum, the locations of glacial refugia, and patterns of subsequent postglacial expansion into northerly latitudes in Britain. We identify populations with unique genetic diversity and examine potential loss of genetic diversity under future climate change scenarios in order to prioritise populations for protection.

2. Methods

2.1 Genetic analyses to map current haplotype diversity

We sampled 146 adults of *E. epiphron* from 13 mountain regions across continental Europe and Britain. European populations (76 adults) were sampled between July - August 2002-2014, populations in England and Scotland (74 adults) were sampled in June-July 2016-2019, and adults preserved in 100% ethanol at -20°C. All relevant fieldwork permissions were obtained. DNA was extracted from 111 individuals with Omega bio-tek E.Z.N.A.® DNA Isolation Kit following the manufacturer's protocol. For each individual, the head and antennae were removed and placed in 1.5 ml tubes with CLT buffer and Proteinase K and homogenised with pellet pestles. A 658-bp fragment of the mitochondrial cytochrome oxidase-I (COI) gene was amplified using the primers LepF (5'-ATTCAACCAATCATAAAGATATTGG-3') and LepR (5'-TAAACTTCTGGATGTCCAAAAATCA-3') (Hajibabaei, Janzen, Burns, Hallwachs, & Hebert, 2006). PCR amplification of individual DNA samples was carried out in 20 µl reactions which included 1.8 µl of template DNA, 1x PCR reaction buffer (Promega), 1.5 mM MgCl₂, 0.2 mM of dNTPs and 1U of *Taq* DNA polymerase (Promega GoTaq®). PCR conditions used the following profile: 94°C for 2 minutes (one cycle), 2 minute at 94°C, 58°C for 45s and 72°C for 1 minute (35 cycles), followed by a final extension step of 75°C for 5 minutes. PCR products were purified and

Sanger sequenced with forward and reverse primers using © Eurofins Scientific PlateSeq service and LightRun Tube service. Chromatograms were checked visually using SeqTrace (Stucky, 2012). Additional COI sequences were obtained from a panel of 39 samples collected in England in June 2016 as a part of a whole genome resequencing project (NERC Highlight project NE/N015797/1). Briefly, the complete mitochondrial genome was assembled for each individual sample using the MitoZ toolkit (Meng et al. 2019) and annotated using the mitos2 webserver (Bernt et al. 2013). Low coverage regions (<10) were masked to avoid introducing low quality SNPs and the COI region was extracted for further analyses.

These 150 sequences along with 65 existing COI sequences from Genbank were combined to create a data set of 215 COI sequences from 13 mountain regions across the species' European range (for sample information see Appendix S1 and map of mountain regions see Appendix S2 in Supporting Information). These sequences were aligned with ClustalX implemented in MEGA-X (Kumar, Stecher, Li, Knyaz, & Tamura, 2018) and the alignment checked by eye and cropped to the same length (649 bp). Haplotypes were identified and genetic diversity measures were determined using DnaSP6 (Rozas et al., 2017). Genetic diversity measures included number of haplotypes (H_n), number of unique haplotypes (H_u), haplotype diversity (H_d , the probability that two randomly sampled alleles are different) and nucleotide diversity (π , the average number of nucleotide differences per site between sequences (Nei, 1987). A TCS network (Templeton, Crandall, & Sing, 1992) of all haplotypes was constructed using PopArt (Leigh & Bryant, 2015). A COI phylogenetic tree was constructed in BEAST (Suchard et al., 2018) of the *Erebia* genus, outgroups and the *E. epiphron* populations. The same methods and COI sequences were used from (Pena et al., 2015) using a log-normal relaxed molecular clock, with a birth-death incomplete speciation model for the randomly generated tree prior, and then an uncorrelated log-normal relaxed molecular clock and all the programs other default settings to model the rate of evolution. The age between *Erebia* and its sister taxa was set at 37.4 ± 2 Myr, (Pena et al., 2015) to estimate age in divergence between *E. epiphron* subpopulations.

2.2 Using species distribution modelling (SDMs) to map current distribution of *E. epiphron*

Current distribution data for *E. epiphron* (50 × 50 km grid resolution) were obtained from the Distribution Atlas of European butterflies (<http://www.ufz.de/european-butterflies/index.php?en=42605>). Current (1970-2000) climate data were downloaded from WorldClim (<http://www.worldclim.org/>) at a resolution of 2.5 arc minutes (~4.5 km grid cell resolution). Climate variables for inclusion in SDMs were selected to reflect climate limitations and extremes of cool-adapted species, which are likely to be most limited by climatic conditions during the coldest and hottest times of the year. We therefore included climate data on annual mean temperature and mean precipitation of the coldest quarter (December to February) and warmest quarter (June to August) of the year (Smith et al., 2013). Spatial autocorrelation was tested using Moran's I in R. The butterfly distribution data were at 50 km grid resolution, but the species is likely to be restricted by local climate conditions in each grid square (Smith et al., 2013). Thus, we included in models only the coldest/warmest and wettest/driest cells (4.5 km resolution) within each 50 km grid, resulting in a total of eight climatic variables being incorporated into our SDMs (see Appendix

S3 in Supporting Information). 50 x 50 km grid cell resolution data are appropriate for our model building to address biogeographic questions at regional scales, because we are interested in changes in the distribution of the study species over long periods of time (i.e. millennia), rather than shorter-term changes at individual sites. This 50km spatial resolution also ensures that the pseudo-absences (i.e. locations where *E. epiphron* is assumed to be absent) are more accurate representations of true absences, because these grids have been visited by butterfly recorders but *E. epiphron* was not recorded as present. In addition, 50km data for presences cover the entire global distribution of *E. epiphron* at this spatial resolution. Butterfly distributions were modelled using an ensemble approach (R package BIOMOD2; (Thuiller, Lafourcade, Engler, & Araujo, 2009), combining outputs from the models; Generalised Linear Models (GLM), Multiple Adaptive Regression Splines (MARS), Maximum Entropy (MAXENT/Phillips), Generalized Additive Model (GAM), Boosted Regression Trees (GBM), Classification Tree Analysis (CTA), Artificial Neural Network (ANN), Surface Range Envelope (SRE), Flexible Discriminant Analysis (FDA) and Random Forest (RF). We used the mean Receiver Operating Characteristic (ROC) value to evaluate each model, with a threshold of ROC > 0.85 for inclusion of models within the ensemble model. We restricted pseudo-absences to locations within a buffer of 250 km around presence data points to avoid placing absences in mountain systems with potentially suitable climate space that are not currently occupied by the species (e.g. Scandinavia) (Akçakaya & Atwood, 1997; Hirzel, Helfer, & Metral, 2001). Models were generated using 70% training data and 30% testing data (Franklin, 2010; Huberty, 1994).

2.3 Hindcasting past distributions and identifying glacial refugia

We incorporated paleoclimate data into our ensemble SDM for the eight climate variables representing the coldest/hottest and driest/wettest locations within each 50 km grid square. Data for climate projections over the last 21,000 years were downloaded from Paleoview ($2.5 \times 2.5^\circ$ (latitude/longitude) grid) (Fordham et al., 2017), and downscaled to match the resolution of the current climate data (2.5 arc minutes), using established methods (Mitasova & Mitas, 1993; Platts, Omeny, & Marchant, 2015; Ramirez-Villegas & Jarvis, 2010). We projected climate suitability for *E. epiphron* every 1,000 years from the LGM to 1,000 years before present, generating 21 outputs, which were each clipped using Eurasian ice sheet data (Hughes, Gyllencreutz, Lohne, Mangerud, & Svendsen, 2016). Long-term climate suitability of 50 km grid squares was calculated by overlaying the 22 output maps and summing the climate-suitability probability values of each grid, and then designating the top 30% of grids with highest probability values as areas of highest long-term climate stability for the study species (Chan, Brown, & Yoder, 2011).

2.4 Projecting future distributions and loss of genetic diversity

Future climate projections for 2070 were obtained from IPCC 5th Assessment Report (Complete Coupled System Model, CCSM4 global climate models) from WorldClim (<http://www.worldclim.org/>; 2.5 arc minutes resolution) for high (RCP 8.5, ~2-3°C warming) and low (RCP 2.6, ~1°C warming) future climate scenarios. Unique haplotypes were assumed to be at risk if all 50 km grid squares in one of the 13 mountain regions were predicted to become climatically unsuitable in the future (based on binary presence or absence threshold probability values from the ensemble SDM output). We set the threshold value as the probability value associated with the low

elevation climatic range edge *E. epiphron* in its current range (low elevation range boundary in England; threshold probability = 0.49). Using this threshold, model probabilities were converted into presence/absence to show grid squares with no change over time (i.e., population persistence), grids predicted to become climatically unsuitable (i.e., extinction), and grids predicted to become climatically suitable (i.e., colonisation). Haplotype risk (H_r) was calculated as the number of unique haplotypes at risk in each of the 13 mountain regions (Figure 1a) due to projected loss of all climatically suitable areas within a region in the future.

3 Results

3.1 Current haplotype diversity across 13 mountain regions in Europe

From our 215 mtDNA samples, we identified 31 mtDNA haplotypes across Europe, including 27 haplotypes unique to a specific mountain region (Figure 1a, Table 1). The high frequency of unique haplotypes across Europe suggests low levels of allele-sharing. There was also high genetic differentiation between populations (AMOVA, $\phi = 0.76$, $p < 0.001$) and the divergence between some of these populations is dated before the last glacial maximum (phylogenetic tree: see Appendix S5 in Supporting Information). The mountain regions containing the highest haplotype diversity include the Pyrenees ($H_d = 0.63$) the western Alps ($H_d = 0.91$) and England ($H_d = 0.64$) (Table 1). The mountain regions containing only unique haplotypes include the Carpathians ($H_u = 2$) and the Tatras ($H_u = 2$). Populations in England ($H_u = 6$) and the western Alps ($H_u = 6$) not only had the highest number of unique haplotypes but also contained some shared haplotypes with other regions (Figure 1a). There are six unique haplotypes in England which diverged from haplotype 8 (Figure 1b), which is present in England, Scotland, Vosges and the western Alps. None of the 6 unique haplotypes in England were found anywhere else, although the presence of the shared haplotype 8 suggests historical allele sharing with the western Alps. Scotland, in addition to the shared haplotype 8, contains one unique haplotype (haplotype 30), which has diverged from haplotype 8 by 1 substitution and shares haplotype 10 with the Apennines (Figure 1). Despite evidence that regions are differentiated, shared haplotypes also provide evidence of historical gene flow across Pyrenees and Cantabrians, and between the Alps and Balkans (Figure 1). The Massif Central population shares one haplotype (haplotype 16) with the Pyrenees and Cantabrian Mountains, and has one unique haplotype (haplotype 29) which diverged from haplotype 16 by one substitution (Figure 1b).

3.2 Modelling the current distribution of *E. epiphron*

Our ensemble SDM was a good fit to the current distribution of *E. epiphron* (95.4% of presences predicted correctly, 76.3% of pseudo-absences predicted correctly (based on the total presence data), ROC = 0.9) (Figure 2a). Areas predicted to be climatically suitable but currently uninhabited by *E. epiphron* include Wales, Scandinavian mountains and eastern Balkans, the latter of which is currently occupied by *Erebia orientalis*. The model rated the minimum temperature of the warmest quarter of the year (June – August) as the most important variable for predicting climate suitability for the species (average importance of this variable across models = 0.73; importance rated from 0-1), probably because this is an important variable in identifying high elevation areas within a 50 km grid square.

3.3 Hindcasting past distributions of *E. epiphron* and identifying areas of long-term survival

Climate suitability in the LGM (21,000 years before present) showed overlap of climatically suitable areas (at 50 km grid resolution) with many locations currently occupied by *E. epiphron*, as well as some southerly locations (Figure 2e). This overlap was confirmed when all 21 SDM outputs for each 1000-year time period up to the present day were combined to show long-term climatic stability since the LGM (Figure 2f). These climate stability maps provided evidence that the locations of glacial refugia were in areas of high topographic variation within the species' current distribution in continental Europe.

3.4 Projecting future distributions and loss of genetic diversity

As expected for a cold-adapted species, SDM outputs from both high and low future climate change scenarios project that many extant *E. epiphron* areas will have reduced climate suitability in the future (38-64% loss of 7,000 km² occupied grids across Europe) (Figure 3, Table 1). The loss of climate suitability is most severe in lower elevation sites, as shown by significant linear regressions between change in probability over time and average elevation of the 50 km grid square (low scenario: $p < 0.001$, $R^2: 0.27$, $F_{150} = 56.51$, high scenario: $p < 0.001$, $R^2: 0.13$, $F_{150} = 22.86$). The mountain regions predicted to experience the greatest reduction in range size are the Vosges (100% loss of grid squares under both scenarios) and Apennines (100% loss of grid squares under both scenarios), followed by the Balkans (75-100% loss), Carpathians (70-100% loss), England (50-100% lost) and Cantabrians (63-81% loss) (Figure 3, Table 1). These range retractions result in the potential loss of 1 haplotype under the low climate change scenario (RCP 2.6); and the total loss of 12 unique haplotypes under the high climate change scenario (Figure 3, Table 1). Many of the haplotypes predicted to be lost are a single substitution from their nearest haplotype, however the haplotypes in the Carpathians are more genetically distinct (Figure 1b). By contrast, range sizes in the Alps and Scotland are projected to remain relatively stable, assuming the species colonises sites at higher elevations that are predicted to become climatically suitable in these regions. Under both scenarios, areas north of Scotland and England become suitable in the future. Although *E. epiphron* does not currently occur in Scandinavia, our models predict that this area will remain stable in climate suitability in the future.

4. Discussion

By using species distribution modelling and mtDNA analyses, we explore the past, present and potential future distributions of genetic diversity in the cold-adapted species *E. epiphron*. We identify high levels of genetic differentiation across Europe, and found evidence of long-term climate suitability in many of these regions since the LGM, which suggests these climatically stable regions were refugial areas of long-term survival by our study species over the last 21,000 years, and potentially longer-term areas of persistence over previous glacial-interglacial cycles. Our study focuses on a single mountain species but our findings are likely to be widely applicable to other mountain species where populations contain unique genetic diversity as a consequence of past climate fluctuations, and which may be at risk under future climate warming. These areas of long-term survival are within topographically heterogeneous landscapes, allowing populations to shift to the foothills during glacial periods. Our analyses also revealed that populations in the Massif Central,

Vosges and Britain are presumed postglacial colonisations (Figure 1, Figure 2f) due to low climate suitability over time, shared haplotypes and the fact that Britain was under an ice sheet during the LGM. Britain was apparently colonised via two different routes, with the Scottish populations likely originating from populations in Vosges/Alps mountain regions due to the high prevalence of shared haplotype 8. By contrast, the English population has high levels of unique genetic diversity, and no evidence that any of the six unique haplotypes are shared with other extant populations (although there is one shared haplotype present), suggesting the English population has separated from the western Alps before the last glacial maximum (given the large number of nucleotide substitutions; Figure 1b), and colonised Britain via a different route, from a cryptic refugium in an area where the study species survived during the glacial period but where it no longer exists. Under future climate change scenarios, we predict 30-56% loss of range size, which equate to 1 unique haplotype to 12 unique haplotypes being at risk of loss under climate scenarios projecting 1 °C and 2-3 °C increases respectfully.

4.1 Limitations

This study has potential limitations, which are inherent in species distribution modelling, especially when projecting into different climates (Buisson, Thuiller, Casajus, Lek, & Grenouillet, 2010). We did not have suitable data to include sampling effort formally into our models and so the areas outside of the current *E. epiphron* distribution are considered ‘pseudo-absences’ rather than ‘true’ absences. However, other butterfly species have been recorded in these squares (Lepidopterists have visited these squares) without recording *E. epiphron* as present, and hence the proportion of false absences in the data is likely to be very low at the spatial (50 km across the whole of Europe) and temporal (accumulation of Lepidoptera records over 3 decades) scales considered here. We consider that our modelling approach robustly describes the bioclimatic conditions occupied by *E. epiphron* at a continental scale (the species’ global distribution). Future work could use sampling effort to account for imperfect species detection, with standardised sampling and occupancy modelling providing additional insight into (especially) within-region distributions and dynamics.

For future projections, the loss of populations and consequently genetic diversity was based on a probability threshold to define butterfly presence or absence. This threshold was based on the probability value for English populations, given that this region represented the lowest elevational range edge for the study species. However, currently realised and fundamental niche characteristics may differ among regions (i.e. thresholds may differ), and hence caution should be taken with our predictions. The difference between using two different thresholds (either the lowest elevation versus a threshold calculated by the Biomod2 program), affects whether or not the entire English and Apennines regions are lost, and hence there is some uncertainty about the level of genetic diversity at risk. Nonetheless, the relatively low probability of future persistence in both of these regions suggests that these populations are at the climatic range limit for the species and therefore at risk. While regional adaptations may differ, we have no evidence that haplotypes are individually adaptive to climate variables and hence we use them as markers of colonisation rather than as adaptive traits. For the same reason, we did not model the specific niches of individual haplotypes when considering the potential future loss of genetic variation (Breiner, Nobis, Bergamini & Guisan et al., 2018). Future

work could use next generation sequencing to further test our hypotheses; and to model specific genetic-climatic relationships in the future (see Bay et al., 2018).

Our analyses suggest that entire mountain regions of the butterfly's distribution could be lost under future climatic change, but it is possible that isolated populations could survive in particular microhabitats, at least temporarily. However, these localised populations may not contain all of the genetic variation currently present in the wider region, and overtime these refugial populations may gradually lose genetic variation and viability (e.g., through inbreeding), and so they may not persist in the longer term due to their isolation (metapopulation failure). A variety of processes may lead to the loss of genetic diversity following isolation, and there can sometimes be a delay in genetic loss following population decline (Kadlec, Vrba, Kepka, Schmitt, & Konvicka, 2010). For example, the sister species of *E. epiphron*, *Erebia orientalis*, is very localised and currently occurs only in the Eastern Balkans and is genetically homogeneous, potentially putting it at risk of inbreeding depression (Hinojosa, Monasterio, Escobes, Dinca, & Vila, 2019). Therefore, our model projections should be seen as representing much longer-term regional-scale expectations, rather than short-term predictions at the local population or microhabitat scale. We believe that our conclusions about the long-term (LGM to present) continental-scale dynamics of *E. epiphron* are robust, and that this knowledge of the past helps frame future risks and provides information for conservation management.

4.2 Long-term survival resulting in unique genetic diversity in cool-adapted species

SDM outputs provide evidence that our exemplar cold-adapted study species occurred in disjunct regions throughout the period from the LGM to the present day, based on the distribution of suitable climate; the genetic data confirm likely separation not only since the LGM, but most probably over much longer periods and successive glacial-interglacial cycles. For mountain species, limited gene flow between the disjunctive parts of their range during glacial and interglacial periods results in divergence and unique haplotypes, unlike lowland European species which colonised northwards from their glacial refugia, and where large parts of the current geographic ranges often share haplotypes (Hewitt, 2004). Only limited areas of postglacial expansions and retractions are evident in *E. epiphron*, and the British populations would be susceptible to extinction if the climate was to return to LGM conditions at some time in the future. Similarly, our SDM outputs suggest that additional populations of *E. epiphron* could have existed further south in southern Europe at the LGM (Figure 2E) but as they no longer exist a northwards translocation of the range might have taken place under interglacial conditions. If cold-adapted species such as *E. epiphron* were more widespread during glacial periods, then the current divergence could be associated with subsequent losses of genetic diversity (e.g., due to selection, or random drift during population bottlenecks), or a failure of our analyses to detect localised or rare haplotype variation. However, this alternative hypothesis seems unlikely because our estimates of times of genetic divergence (phylogenetic tree: see Appendix S5) imply that most splits occurred before the LGM. However, other divergence dates between *E. epiphron* and *E. orientalis* have been reported (e.g. 1.53 (± 0.65) Mya (Hinojosa et al., 2019)). However they still reported strong mtDNA divergence and long term separation (Hinojosa et al., 2019) and therefore different assumptions of divergence dates do not impact the interpretation of our

results. Hence we conclude that populations of *E. epiphron* survived as allopatric populations in mainland Europe during the LGM, with postglacial colonisations from these regions into the Massif Central, Vosges, Scotland and England.

High genetic differentiation is observed among populations of other mountain *Erebia* species, supporting the hypothesis that they also survived as allopatric populations during the LGM (Haubrich & Schmitt, 2007; Louy, Habel, Abadjiev, et al., 2014; Louy, Habel, Ulrich, & Schmitt, 2014; Martin, Gilles, Lortscher, & Descimon, 2002; Schmitt et al., 2014; Schmitt, Louy, Zimmermann, & Habel, 2016; Schmitt & Seitz, 2001). LGM separation of populations has also been identified in mountain plants and other invertebrates (Bettin, Cornejo, Edwards, & Holderegger, 2007; Huck, Budel, & Schmitt, 2012; Margraf, Verdon, Rahier, & Naisbit, 2007; Pauls, Lumbsch, & Haase, 2006). The numbers of glacial-interglacial cycles over which populations have remained disjunct remains unclear, but some studies have indicated divergence dates covering several glacial-interglacial cycles or even predating the Pleistocene (Hewitt, 2000). The reality is likely to be more complex with areas of persistent separation, but with occasional links between them (i.e. rare gene flow or brief periods of connection), as indicated by the distributions and relatedness of haplotypes in Figure 1.

4.3 Unique haplotypes in populations derived from northern cryptic refugia

Following the LGM, the ice retreated in northern Europe and many species colonised northwards, for example via the land bridge between continental Europe and Britain, which was present until sea level rise ~7,000 years before present (Sturt, Garrow, & Bradley, 2013). The locations of southerly glacial refugia, which are thought to be the main sources of colonisations, have been debated extensively, with proposed glacial refugia in the Iberian Peninsula, Italy and the Balkans (Hewitt, 2000) and this has recently been reinforced in European butterflies (Dapporto et al., 2019). However, there is also evidence for more northern cryptic refugia based on fossil, pollen and genetic evidence (Birks & Willis, 2008; Provan and Bennett, 2008; Stewart and Lister, 2001), where species apparently persisted at higher latitudes in sheltered locations with suitable microclimates (Stewart, Lister, Barnes, & Dalen, 2010). However, most cryptic refugia described to date have been for relatively warm-adapted species. Here, we present evidence for the existence of northern cryptic population(s) for cold-adapted species during the LGM, based on high unique genetic diversity of the present-day *E. epiphron* populations in England, an area that was beneath an ice sheet at the LGM (Hughes et al., 2016). The high genetic uniqueness of populations in England, together with a single shared haplotype with Scotland/Vosges/Alps (haplotype 8; Figure 1b), is consistent with northern colonisations from the Alps, but distinct separate colonisation of Britain via two routes, although there are alternative explanations. For example, the 6 unique haplotypes in populations in England might occur elsewhere but were not detected in this study. Alternatively, the six unique haplotypes identified in England could have diverged from the shared haplotype in Scotland, Vosges and Alps populations (haplotype 8; Figure 1b) since the LGM, although this seems highly unlikely given the short time period for one to three mutations to occur (Figure 1b). It is possible that these LGM populations were situated on land that is currently below sea level, at an edge of the glacier, or in sheltered low elevation microclimates on land. Multiple colonisation events have also been shown in other taxa in the UK (Piertney et al., 2005), and the locations of cryptic refugia during the LGM are assumed to be ice free

areas in southern England (Bocherens et al., 1995, Lister, 1984), northern Scotland (Bennett, 1984) and southern Ireland (Montgomery et al., 2014). Evidence for cryptic refugia for insects in Britain also comes from cold-adapted beetles (see Appendix S6 in Supporting Information; (Buckland & Buckland, 2006)), which currently have mountain or northern distributions in the UK, but were found as sub-fossil remains in southern England 18,000-26,000 years BP, providing evidence of cold-adapted insects surviving in ice-free locations in Britain in the LGM. It is, therefore, possible that the current population of *E. epiphron* in England survived elsewhere in Britain during the LGM as populations which no longer exists.

4.4 Future loss of unique genetic diversity in cold-adapted species

High levels of genetic diversity are important in relation to the capacity for populations and species to adapt to changing environmental conditions, including climate change (Balint et al., 2011; Hoffmann & Sgro, 2011). Cold-adapted species that have been shaped by diversification across mountain systems during the Pleistocene contain high levels of genetic diversity and unique populations, and are under threat from climate warming. Populations with unique genetic diversity may have evolved independently to be adapted to their local environment (Weeks, Stoklosa, & Hoffmann, 2016) and thus may be particularly vulnerable to future climatic changes. Our SDMs project loss of suitable climate for *E. epiphron* in many locations in Europe, especially in regions with predominantly low elevation populations and few opportunities to shift uphill to high elevation, which could result in loss of genetic diversity. However, our projections of range retraction do not take into account any potential of populations to adapt to warmer temperatures *in situ* (Franks & Hoffmann, 2012). Future loss of genetic diversity has also been predicted in other species (Alsos et al., 2012; Beatty & Provan, 2011; Yannic et al., 2014), and rates of loss of genetic diversity in wild populations since the industrial revolution (Leigh, Hendry, Vázquez-Domínguez, & Friesen, 2019) are consistent with our projections.

4.5 Conservation interventions to mitigate climate-driven genetic erosion

Conservation management and adaptation could protect cold-adapted populations and safeguard unique genetic diversity from climate change (Mawdsley, O'Malley, & Ojima, 2009). Options include translocation or assisted colonisation to areas that have, or are predicted to have, suitable climate and habitat in the future (Hoegh-Guldberg et al., 2008). Translocations are a controversial topic due to the fear that translocated species may become 'invasive' in their new ranges, posing threats to ecosystems including disturbance, disrupting ecological interactions, disease spread, competition and extinctions (Ricciardi & Simberloff, 2009). However, others argue that the arrival of new species is typical of ecosystem changes in the Anthropocene, and that translocations mirror colonisations occurring as a consequence of current environmental change (Thomas, 2011). Translocations of *E. epiphron* and other butterflies into unoccupied but climatically-suitable areas have been successful (Cizek, Bakesova, Kuras, Benes, & Konvicka, 2003; Willis et al., 2009), and cold-adapted insects may represent good targets for translocations given that the climate is rapidly deteriorating for them in many parts of their range, and they may find it difficult to colonise new areas across inhospitable landscapes (Thomas, 2011). For *E. epiphron*, our SDMs reveal areas in Scandinavia to be climatically suitable, although the species does not occur there, and climate is predicted to increase in suitability in

future in Scandinavia for *E. epiphron* (Fig 3) and for other *Erebia* species (Settele et al., 2008). However, although Scandinavia may have suitable climate, it may not have the required habitat for *E. epiphron*. Local translocations within mountain systems that are currently occupied by *E. epiphron* could also be implemented, for example moving individuals to areas of colder climate at higher elevation, or neighbouring mountains which are too isolated for the species to colonise naturally. However there may be very few areas of unoccupied but climatically-suitable habitats within some mountain systems occupied by *E. epiphron*, particularly if the species already occurs at high elevations in these regions. Future work could include finer scale country specific SDMs with additional land use and genetic data on habitat availability could be used to locate areas for potential translocations.

As well as translocating individuals to new sites, it might be possible to consider translocating genes or ‘genetic rescue’ by moving individuals among existing populations. Not only might this conserve unique genetic diversity at risk from local extinction of populations, but might increase the adaptive capacity of populations by increasing their genetic diversity (Aitken & Whitlock, 2013). This could involve moving warm-adapted individuals into cooler populations to increase their adaptive capacity as the climate warms (Weeks et al., 2011). However, moving locally-adapted populations may result in outbreeding depression and maladaptation, negatively impacting populations (Weeks et al., 2011), although some genetic rescue interventions have resulted in increases in populations, and alleles associated with local adaptation were not lost following gene flow (Fitzpatrick et al., 2020). Genetic conservation interventions for insects, and specifically butterflies, has been rarely implemented, although increasing habitat connectivity has led to genetic rescue of populations (Jangjoo, Matter, Roland, & Keyghobadi, 2016) and genetic data have been used to inform on reintroductions (Dinca et al., 2018). There is no evidence of attempted genetic rescue via translocations of butterflies, although translocating individuals is a genetic conservation strategy which may be important in ensuring future survival and adaptability of populations under climate change. As with translocations, these conservation options may also be controversial, but could remove the need for on-going intervention and management at sites with declining populations (Weeks et al., 2011). We recommend that before the implementation of any climate adaptation strategy, populations are closely monitored to determine if populations are retracting and likely to become extinct in areas that are becoming too warm for the species. In addition, individual species’ assessments are required to assess the genetic diversity of populations and any local adaptation, which would determine the most appropriate conservation strategy.

5. Conclusions

The genetic diversification of cool-adapted mountain species, as demonstrated in our study species *E. epiphron*, has been shaped by Pleistocene glaciations, the locations of long-term survival of populations, and colonisation patterns after the LGM, resulting in unique genetic diversity in isolated populations. Mountain and cold-adapted species are vulnerable to future climate warming, and we predict *E. epiphron* will lose 30-55% of its range in the future, especially at low elevations. The uniqueness of genetic diversity contained in these populations could be at risk depending on the severity of future climate change. Conservation strategies such as translocation could ensure the

survival of these cold-adapted species, but more research is needed on the likely effectiveness of such approaches.

References

- Aitken, S. N., & Whitlock, M. C. (2013). Assisted Gene Flow to Facilitate Local Adaptation to Climate Change. *Annual Review of Ecology, Evolution, and Systematics*, 44, 367-388. doi:10.1146/annurev-ecolsys-110512-135747
- Akcakaya, H. R., & Atwood, J. L. (1997). A habitat based metapopulation model of the California gnatcatcher. *Conservation Biology*, 11(2), 422-434. doi:10.1046/j.1523-1739.1997.96164.x
- Alsos, I. G., Ehrich, D., Thuiller, W., Eidesen, P. B., Tribsch, A., Schonswetter, P., . . . Brochmann, C. (2012). Genetic consequences of climate change for northern plants. *Proceedings of the Royal Society B-Biological Sciences*, 279(1735), 2042-2051. doi:10.1098/rspb.2011.2363
- Bay, R. A., Harrigan, R. J., Le Underwood, V., Gibbs, H. L., Smith, T. B., & Ruegg, K. (2018). Genomic signals of selection predict climate-driven population declines in a migratory bird. *Science*, 359(6371). doi:10.1126/science.aan4380
- Balint, M., Domisch, S., Engelhardt, C. H. M., Haase, P., Lehrian, S., Sauer, J., . . . Nowak, C. (2011). Cryptic biodiversity loss linked to global climate change. *Nature Climate Change*, 1(6), 313-318. doi:10.1038/nclimate1191
- Beatty, G. E., & Provan, J. (2011). Comparative phylogeography of two related plant species with overlapping ranges in Europe, and the potential effects of climate change on their intraspecific genetic diversity. *Bmc Evolutionary Biology*, 11(29). doi:10.1186/1471-2148-11-29
- Bernt, M., Donath, A., Juhling, F., Externbrink, F., Florentz, C., Fritzsche, G., Putz, J., Middendorf, M., & Stadler, P. F. (2013) MITOS: Improved de novo metazoan mitochondrial genome annotation, *Molecular Phylogenetics and Evolution*, 69(2), 313-319 doi:10.1016/j.ympev.2012.08.023
- Bettin, O., Cornejo, C., Edwards, P. J., & Holderegger, R. (2007). Phylogeography of the high alpine plant *Senecio halleri* (Asteraceae) in the European Alps: in situ glacial survival with postglacial stepwise dispersal into peripheral areas. *Molecular Ecology*, 16(12), 2517-2524. doi:10.1111/j.1365-294X.2007.03273.x
- Birks, H. J. B., & Willis, K. J. (2008). Alpines, trees, and refugia in Europe. *Plant Ecology & Diversity*, 1(2), 147-160. doi:10.1080/17550870802349146
- Botkin, D. B., Saxe, H., Araujo, M. B., Betts, R., Bradshaw, R. H. W., Cedhagen, T., . . . Stockwell, D. R. B. (2007). Forecasting the effects of global warming on biodiversity. *Bioscience*, 57(3), 227-236. doi:10.1641/b570306
- Breiner, F. T., Nobis, M. P., Bergamini, A., & Guisan, A. (2018). Optimizing ensembles of small models for predicting the distribution of species with few occurrences. *Methods in Ecology and Evolution*, 9(4), 802-808.
- Buisson, L., Thuiller, W., Casajus, N., Lek, S., & Grenouillet, G. (2010). Uncertainty in ensemble forecasting of species distribution. *Global Change Biology*, 16(4), 1145-1157. doi:10.1111/j.1365-2486.2009.02000.x
- Buckland, P. I., & Buckland, P. C. (2006). *BugsCEP Coleopteran Ecology Package*. IGBP PAGES/World Data Center for Paleoclimatology Data Contribution Series, NOAA/NCDC Paleoclimatology Program, Boulder CO, USA. <http://www.bugscep.com>.
- Chan, L. M., Brown, J. L., & Yoder, A. D. (2011). Integrating statistical genetic and geospatial methods brings new power to phylogeography. *Molecular Phylogenetics and Evolution*, 59(2), 523-537. doi:10.1016/j.ympev.2011.01.020
- Cizek, O., Bakesova, A., Kuras, T., Benes, J., & Konvicka, M. (2003). Vacant niche in alpine habitat: the case of an introduced population of the butterfly *Erebia epiphron* in the Krkonose Mountains. *Acta Oecologica-International Journal of Ecology*, 24(1), 15-23. doi:10.1016/s1146-609x(02)00004-8

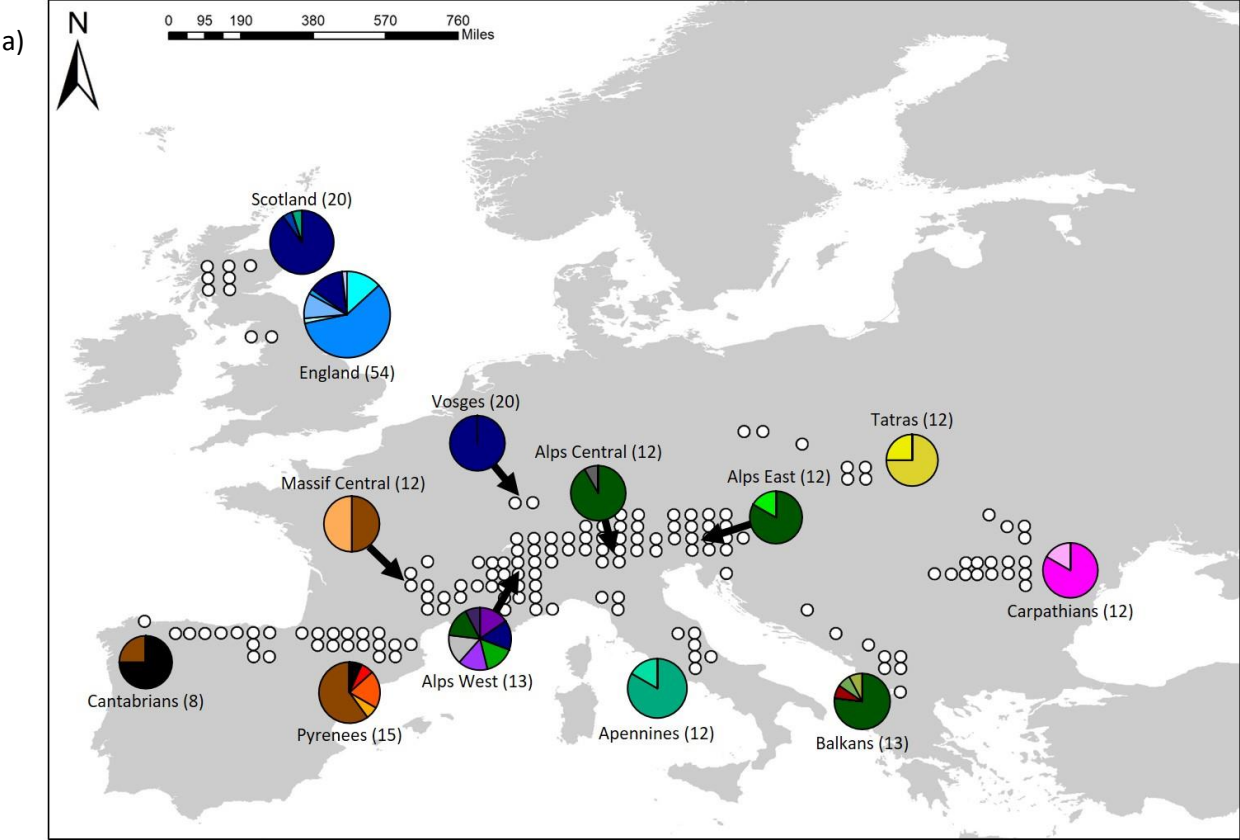
- Crowley, T. J., & North, G., R. (1991). *Palaeoclimatology*. Oxford University Press, Oxford.
- Dapporto, L., Cini, A., Voda, R., Dinca, V., Wiemers, M., Menchetti, M., . . . Vila, R. (2019). Integrating three comprehensive data sets shows that mitochondrial DNA variation is linked to species traits and paleogeographic events in European butterflies. *Molecular Ecology Resources*, 19(6), 1623-1636. doi:10.1111/1755-0998.13059
- Deutsch, C. A., Tewksbury, J. J., Huey, R. B., Sheldon, K. S., Ghalambor, C. K., Haak, D. C., & Martin, P. R. (2008). Impacts of climate warming on terrestrial ectotherms across latitude. *Proceedings of the National Academy of Sciences of the United States of America*, 105(18), 6668-6672. doi:10.1073/pnas.0709472105
- Dinca, V., Balint, Z., Voda, R., Dapporto, L., Hebert, P. D. N., & Vila, R. (2018). Use of genetic, climatic, and microbiological data to inform reintroduction of a regionally extinct butterfly. *Conservation Biology*, 32(4), 828-837. doi:10.1111/cobi.13111
- Elsen, P. R., & Tingley, M. W. (2015). Global mountain topography and the fate of montane species under climate change. *Nature Climate Change*, 5(8), 772-776. doi:10.1038/nclimate2656
- Fitzpatrick, S. W., Bradburd, G. S., Kremer, C. T., Salerno, P. E., Angeloni, L. M., & Funk, C. W. (2020). Genomic and fitness consequences of genetic rescue in wild populations. *Current Biology*, 30, 1-6. doi:10.1016/j.cub.2019.11.062
- Fordham, D. A., Saltre, F., Haythorne, S., Wigley, T. M. L., Otto-Bliesner, B. L., Chan, K. C., & Brook, B. W. (2017). PaleoView: a tool for generating continuous climate projections spanning the last 21 000 years at regional and global scales. *Ecography*, 40(11), 1348-1358. doi:10.1111/ecog.03031
- Franco, A. M. A., Hill, J. K., Kitschke, C., Collingham, Y. C., Roy, D. B., Fox, R., . . . Thomas, C. D. (2006). Impacts of climate warming and habitat loss on extinctions at species' low-latitude range boundaries. *Global Change Biology*, 12(8), 1545-1553. doi:10.1111/j.1365-2486.2006.01180.x
- Franklin, J. (2010). *Mapping Species Distributions, Spatial Inference and Prediction*. Cambridge University Press.
- Franks, S. J., & Hoffmann, A. A. (2012). Genetics of Climate Change Adaptation. *Annual Review of Genetics*, 46, 185-208. doi:10.1146/annurev-genet-110711-155511
- Guo, Y. L., Li, X., Zhao, Z. F., Wei, H. Y., Gao, B., & Gu, W. (2017). Prediction of the potential geographic distribution of the ectomycorrhizal mushroom *Tricholoma matsutake* under multiple climate change scenarios. *Scientific Reports*, 7(46221). doi:10.1038/srep46221
- Hajibabaei, M., Janzen, D. H., Burns, J. M., Hallwachs, W., & Hebert, P. D. N. (2006). DNA barcodes distinguish species of tropical Lepidoptera. *Proceedings of the National Academy of Sciences of the United States of America*, 103(4), 968-971. doi:10.1073/pnas.0510466103
- Haubrich, K., & Schmitt, T. (2007). Cryptic differentiation in alpine-endemic, high-altitude butterflies reveals down-slope glacial refugia. *Molecular Ecology*, 16(17), 3643-3658. doi:10.1111/j.1365-294X.2007.03424.x
- Hewitt, G. (2000). The genetic legacy of the Quaternary ice ages. *Nature*, 405(6789), 907-913. doi:10.1038/35016000
- Hewitt, G. M. (2004). Genetic consequences of climatic oscillations in the Quaternary, *Philosophical Transactions of the Royal Society of London Series B-Biological Sciences*, 359(1442), 183-195. doi:10.1098/rstb.2003.1388
- Hinojosa, J. C., Monasterio, Y., Escobes, R., Dinca, V., & Vila, R. (2019). *Erebia epiphron* and *Erebia orientalis*: sibling butterfly species with contrasting histories. *Biological Journal of the Linnean Society*, 126(2), 338-348. doi:10.1093/biolinnean/bly182
- Hirzel, A. H., Helfer, V., & Metral, F. (2001). Assessing habitat-suitability models with a virtual species. *Ecological Modelling*, 145, 111-121. doi:10.1016/s0304-3800(01)00396-9
- Hoegh-Guldberg, O., Hughes, L., McIntyre, S., Lindenmayer, D. B., Parmesan, C., Possingham, H. P., & Thomas, C. D. (2008). Assisted colonization and rapid climate change. *Science*, 321(5887), 345-346. doi:10.1126/science.1157897
- Hoffmann, A. A., & Sgro, C. M. (2011). Climate change and evolutionary adaptation. *Nature*, 470(7335), 479-485. doi:10.1038/nature09670
- Huberty, C., J. (1994). *Applied Discriminant Analysis*. New York, USA: Wiley International science.

- Huck, S., Budel, B., & Schmitt, T. (2012). Ice-age isolation, postglacial hybridization and recent population bottlenecks shape the genetic structure of *Meum athamanticum* in Central Europe. *Flora*, 207(6), 399-407. doi:10.1016/j.flora.2012.03.005
- Hughes, A. L. C., Gyllencreutz, R., Lohne, O. S., Mangerud, J., & Svendsen, J. I. (2016). The last Eurasian ice sheets - a chronological database and time-slice reconstruction, DATED-1. *Boreas*, 45(1), 1-45. doi:10.1111/bor.12142
- Jangjoo, M., Matter, S. F., Roland, J., & Keyghobadi, N. (2016). Connectivity rescues genetic diversity after a demographic bottleneck in a butterfly population network. *Proceedings of the National Academy of Sciences of the United States of America*, 113(39), 10914-10919. doi:10.1073/pnas.1600865113
- Kadlec, T., Vrba, P., Kepka, P., Schmitt, T., & Konvicka, M. (2010). Tracking the decline of the once-common butterfly: delayed oviposition, demography and population genetics in the hermit *Chazara briseis*. *Animal Conservation*, 13(2), 172-183. doi:10.1111/j.1469-1795.2009.00318.x
- Keppel, G., Van Niel, K. P., Wardell-Johnson, G. W., Yates, C. J., Byrne, M., Mucina, L., . . . Franklin, S. E. (2012). Refugia: identifying and understanding safe havens for biodiversity under climate change. *Global Ecology and Biogeography*, 21(4), 393-404. doi:10.1111/j.1466-8238.2011.00686.x
- Kumar, S., Stecher, G., Li, M., Knyaz, C., & Tamura, K. (2018). MEGA X: Molecular Evolutionary Genetics Analysis across Computing Platforms. *Molecular Biology and Evolution*, 35(6), 1547-1549. doi:10.1093/molbev/msy096
- Leigh, D., M., Hendry, A. P., Vázquez-Domínguez, E. & Friesen, V. L. (2019). Estimated six per cent loss of genetic variation in wild populations since the industrial revolution. *Evolutionary Applications*, 12(8), 1-8.
- Leigh, J. W., & Bryant, D. (2015). POPART: full-feature software for haplotype network construction. *Methods in Ecology and Evolution*, 6(9), 1110-1116. doi:10.1111/2041-210x.12410
- Louy, D., Habel, J. C., Abadjiev, S., Rakosy, L., Varga, Z., Rodder, D., & Schmitt, T. (2014). Molecules and models indicate diverging evolutionary effects from parallel altitudinal range shifts in two mountain Ringlet butterflies. *Biological Journal of the Linnean Society*, 112(3), 569-583. doi:10.1111/bij.12240
- Louy, D., Habel, J. C., Ulrich, W., & Schmitt, T. (2014). Out of the Alps: The Biogeography of a Disjunctly Distributed Mountain Butterfly, the Almond-Eyed Ringlet *Erebia albertanus* (Lepidoptera, Satyrinae). *Journal of Heredity*, 105(1), 28-38. doi:10.1093/jhered/est081
- Luquet, E., Morch, P. R., Cortazar-Chinarro, M., Meyer-Lucht, Y., Høglund, J., & Laurila, A. (2019). Post-glacial colonization routes coincide with a life-history breakpoint along a latitudinal gradient. *Journal of Evolutionary Biology*, 32(4), 356-368. doi:10.1111/jeb.13419
- Margraf, N., Verdon, A., Rahier, M., & Naisbit, R. E. (2007). Glacial survival and local adaptation in an alpine leaf beetle. *Molecular Ecology*, 16(11), 2333-2343. doi:10.1111/j.1365-294X.2007.03318.x
- Martin, J. F., Gilles, A., Lortscher, M., & Descimon, H. (2002). Phylogenetics and differentiation among the western taxa of the *Erebia tyndarus* group (Lepidoptera : Nymphalidae). *Biological Journal of the Linnean Society*, 75(3), 319-332.
- Mawdsley, J. R., O'Malley, R., & Ojima, D. S. (2009). A Review of Climate-Change Adaptation Strategies for Wildlife Management and Biodiversity Conservation. *Conservation Biology*, 23(5), 1080-1089. doi:10.1111/j.1523-1739.2009.01264.x
- McCallum, K. P., Guerin, G. R., Breed, M. F., & Lowe, A. J. (2014). Combining population genetics, species distribution modelling and field assessments to understand a species vulnerability to climate change. *Austral Ecology*, 39(1), 17-28. doi:10.1111/aec.12041
- Meng, G., Li, Y., Yang, C., & Liu, S. (2019) MitoZ: a toolkit for animal mitochondrial genome assembly, annotation and visualization. *Nucleic Acids Research*, 47(11). doi:10.1093/nar/gkz173
- Mitasova, H., & Mitas, L. (1993). Interpolation by regularized spline with tension .1. theory and implementation. *Mathematical Geology*, 25(6), 641-655. doi:10.1007/bf00893171

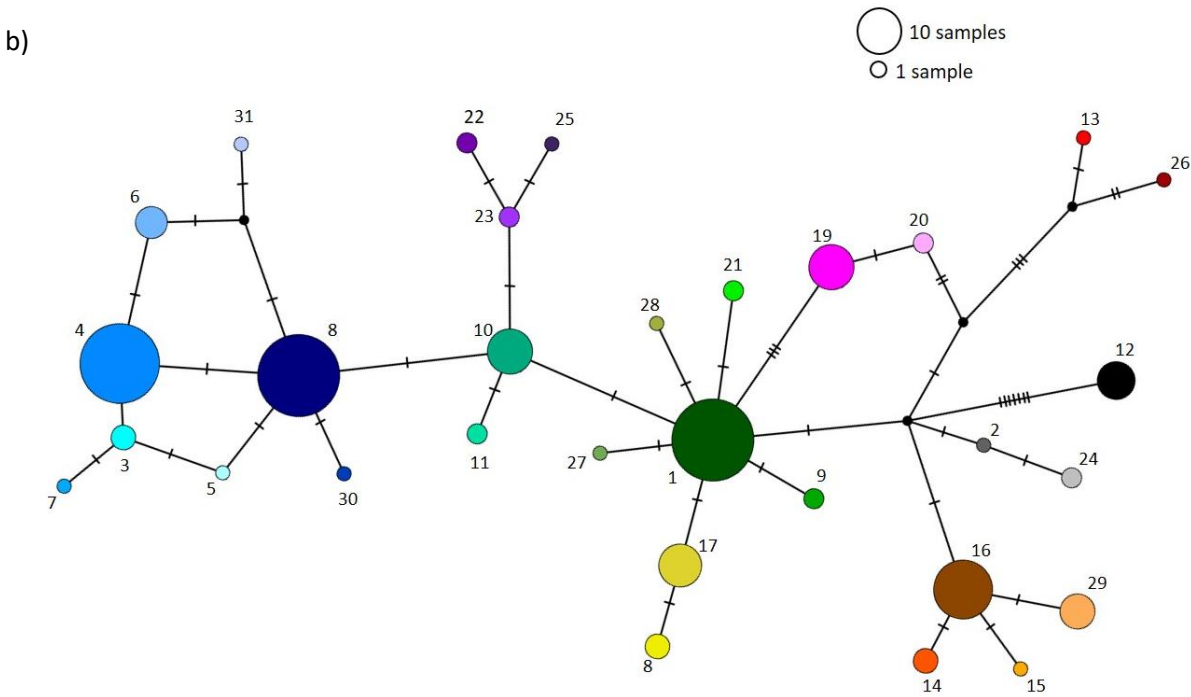
- Morelli, T. L., Daly, C., Dobrowski, S. Z., Dulen, D. M., Ebersole, J. L., Jackson, S. T., . . . Beissinger, S. R. (2016). Managing Climate Change Refugia for Climate Adaptation. *Plos One*, 11(8). doi:10.1371/journal.pone.0159909
- Nei, M. (1987). *Molecular Evolutionary Genetics*. New York: Columbia Univ. Press
- Pauls, S. U., Lumbsch, H. T., & Haase, P. (2006). Phylogeography of the montane caddisfly *Drusus discolor*: evidence for multiple refugia and periglacial survival. *Molecular Ecology*, 15(8), 2153-2169. doi:10.1111/j.1365-294X.2006.02916.x
- Piertney, S. B., Stewart, W. A., Lambin, X., Telfer, S., Aars, J., & Dallas, J. F. (2005). Phylogeographic structure and postglacial evolutionary history of water voles (*Arvicola terrestris*) in the United Kingdom. *Molecular Ecology*, 14(5), 1435-1444. doi:10.1111/j.1365-294X.2005.02496.x
- Platts, P. J., Omeny, P. A., & Marchant, R. (2015). AFRICLIM: high-resolution climate projections for ecological applications in Africa. *African Journal of Ecology*, 53(1), 103-108. doi:10.1111/aje.12180
- Provan, J., & Bennett, K. D. (2008). Phylogeographic insights into cryptic glacial refugia. *Trends in Ecology & Evolution*, 23(10), 564-571. doi:10.1016/j.tree.2008.06.010
- Ramirez-Villegas, J., & Jarvis, A. (2010). *Downscaling Global Circulation Model Outputs: The Delta Method*. CIAT Decision and Policy Analysis Working Paper, no. 1. International Center for Tropical Agriculture (CIAT)
- Ray, N., & Adams, J. (2001). GIS-based vegetation map of the world at the Last Glacial Maximum (25,000-15,000 BP). *Internet Archaeology*, 11(1).
- Ricciardi, A., & Simberloff, D. (2009). Assisted colonization is not aviable conservation strategy. *Trends Ecol. Evo*, 24(5), 248-253. doi:10.1016/j.tree.2008.12.006
- Rozas, J., Ferrer-Mata, A., Sanchez-DelBarrio, J. C., Guirao-Rico, S., Librado, P., Ramos-Onsins, S. E., & Sanchez-Gracia, A. (2017). DnaSP 6: DNA Sequence Polymorphism Analysis of Large Data Sets. *Molecular Biology and Evolution*, 34(12), 3299-3302. doi:10.1093/molbev/msx248
- Santer, B. D., Bonfils, C. J. W., Fu, Q., Fyfe, J. C., Hegerl, G. C., Mears, C., . . . Zou, C. Z. (2019). Celebrating the anniversary of three key events in climate change science. *Nature Climate Change*, 9(3), 180-182. doi:10.1038/s41558-019-0424-x
- Schmitt, T. (2009). Biogeographical and evolutionary importance of the European high mountain systems. *Frontiers in Zoology*, 6(9). doi:10.1186/1742-9994-6-9
- Schmitt, T., Habel, J. C., Rodder, D., & Louy, D. (2014). Effects of recent and past climatic shifts on the genetic structure of the high mountain Yellow-spotted ringlet butterfly *Erebia manto* (Lepidoptera, Satyrinae): a conservation problem. *Global Change Biology*, 20(7), 2045-2061. doi:10.1111/gcb.12462
- Schmitt, T., Hewitt, G. M., & Muller, P. (2006). Disjunct distributions during glacial and interglacial periods in mountain butterflies: *Erebia epiphron* as an example. *Journal of Evolutionary Biology*, 19(1), 108-113. doi:10.1111/j.1420-9101.2005.00980.x
- Schmitt, T., Louy, D., Zimmermann, E., & Habel, J. C. (2016). Species radiation in the Alps: multiple range shifts caused diversification in Ringlet butterflies in the European high mountains. *Organisms Diversity & Evolution*, 16(4), 791-808. doi:10.1007/s13127-016-0282-6
- Schmitt, T., & Seitz, A. (2001). Intraspecific allozymatic differentiation reveals the glacial refugia and the postglacial expansions of European *Erebia medusa* (Lepidoptera : Nymphalidae). *Biological Journal of the Linnean Society*, 74(4), 429-458. doi:10.1006/bjil.2001.0584
- Schmitt, T., & Varga, Z. (2012). Extra-Mediterranean refugia: The rule and not the exception? *Front Zool*, 9(22).
- Settele, J., Kudrna, O., Harpke, A., Kühn, I., van Swaay, C., Verovnik, R., . . . Schweiger, O. (2008). *Climatic Risk Atlas of European Butterflies*. Sofia, Moscow: Pensoft
- Smith, S. E., Gregory, R. D., Anderson, B. J., & Thomas, C. D. (2013). The past, present and potential future distributions of cold-adapted bird species. *Diversity and Distributions*, 19(3), 352-362. doi:10.1111/ddi.12025
- Stewart, J. R., & Lister, A. M. (2001). Cryptic northern refugia and the origins of the modern biota. *Trends in Ecology & Evolution*, 16(11), 608-613. doi:10.1016/s0169-5347(01)02338-2

- Stewart, J. R., Lister, A. M., Barnes, I., & Dalen, L. (2010). Refugia revisited: individualistic responses of species in space and time. *Proceedings of the Royal Society B-Biological Sciences*, 277(1682), 661-671. doi:10.1098/rspb.2009.1272
- Stucky, B., J. (2012). SeqTrace: A Graphical Tool for Rapidly Processing DNA Sequencing Chromatograms. *Journal of Biomolecular Techniques*, 23(3), 90-93.
- Sturt, F., Garrow, D., & Bradley, S. (2013). New models of North West European Holocene palaeogeography and inundation. *Journal of Archaeological Science*, 40(11), 3963-3976. doi:10.1016/j.jas.2013.05.023
- Suchard, M. A., Lemey, P., Baele, G., Ayres, D. L., Drummond, A. J., & Rambaut, A. (2018). Bayesian phylogenetic and phylodynamic data integration using BEAST 1.10. *Virus Evolution*, 4(1). doi:10.1093/ve/vey016
- Templeton, A. R., Crandall, K. A., & Sing, C. F. (1992). A cladistic analysis of phenotypic associations with haplotypes inferred from restriction endonuclease mapping and DNA sequence data. III. Cladogram estimation. *Genetics*, 132(2), 619-633.
- Thomas, C. D. (2010). Climate, climate change and range boundaries. *Diversity and Distributions*, 16(3), 488-495. doi:10.1111/j.1472-4642.2010.00642.x
- Thomas, C. D. (2011). Translocation of species, climate change, and the end of trying to recreate past ecological communities. *Trends in Ecology & Evolution*, 26(5), 216-221. doi:10.1016/j.tree.2011.02.006
- Thuiller, W., Lafourcade, B., Engler, R., & Araujo, M. B. (2009). BIOMOD - a platform for ensemble forecasting of species distributions. *Ecography*, 32(3), 369-373. doi:10.1111/j.1600-0587.2008.05742.x
- Urban, M. C. (2015). Accelerating extinction risk from climate change. *Science*, 348(6234), 571-573. doi:10.1126/science.aaa4984
- Weeks, A. R., Sgro, C. M., Young, A. G., Frankham, R., Mitchell, N. J., Miller, K. A., . . . Hoffmann, A. A. (2011). Assessing the benefits and risks of translocations in changing environments: a genetic perspective. *Evolutionary Applications*, 4(6), 709-725. doi:10.1111/j.1752-4571.2011.00192.x
- Weeks, A. R., Stoklosa, J., & Hoffmann, A. A. (2016). Conservation of genetic uniqueness of populations may increase extinction likelihood of endangered species: the case of Australian mammals. *Frontiers in Zoology*, 13(31). doi:10.1186/s12983-016-0163-z
- Willis, S. G., Hill, J. K., Thomas, C. D., Roy, D. B., Fox, R., Blakeley, D. S., & Huntley, B. (2009). Assisted colonization in a changing climate: a test-study using two UK butterflies. *Conservation Letters*, 2(1), 45-51. doi:10.1111/j.1755-263X.2008.00043.x
- Wilson, R. J., Gutierrez, D., Gutierrez, J., & Monserrat, V. J. (2007). An elevational shift in butterfly species richness and composition accompanying recent climate change. *Global Change Biology*, 13(9), 1873-1887. doi:10.1111/j.1365-2486.2007.01418.x
- Wroblewska, A., & Mirski, P. (2018). From past to future: impact of climate change on range shifts and genetic diversity patterns of circumboreal plants. *Regional Environmental Change*, 18(2), 409-424. doi:10.1007/s10113-017-1208-3
- Yannic, G., Pellissier, L., Ortego, J., Lecomte, N., Couturier, S., Cuyler, C., . . . Cote, S. D. (2014). Genetic diversity in caribou linked to past and future climate change. *Nature Climate Change*, 4(2), 132-137. doi:10.1038/nclimate2074

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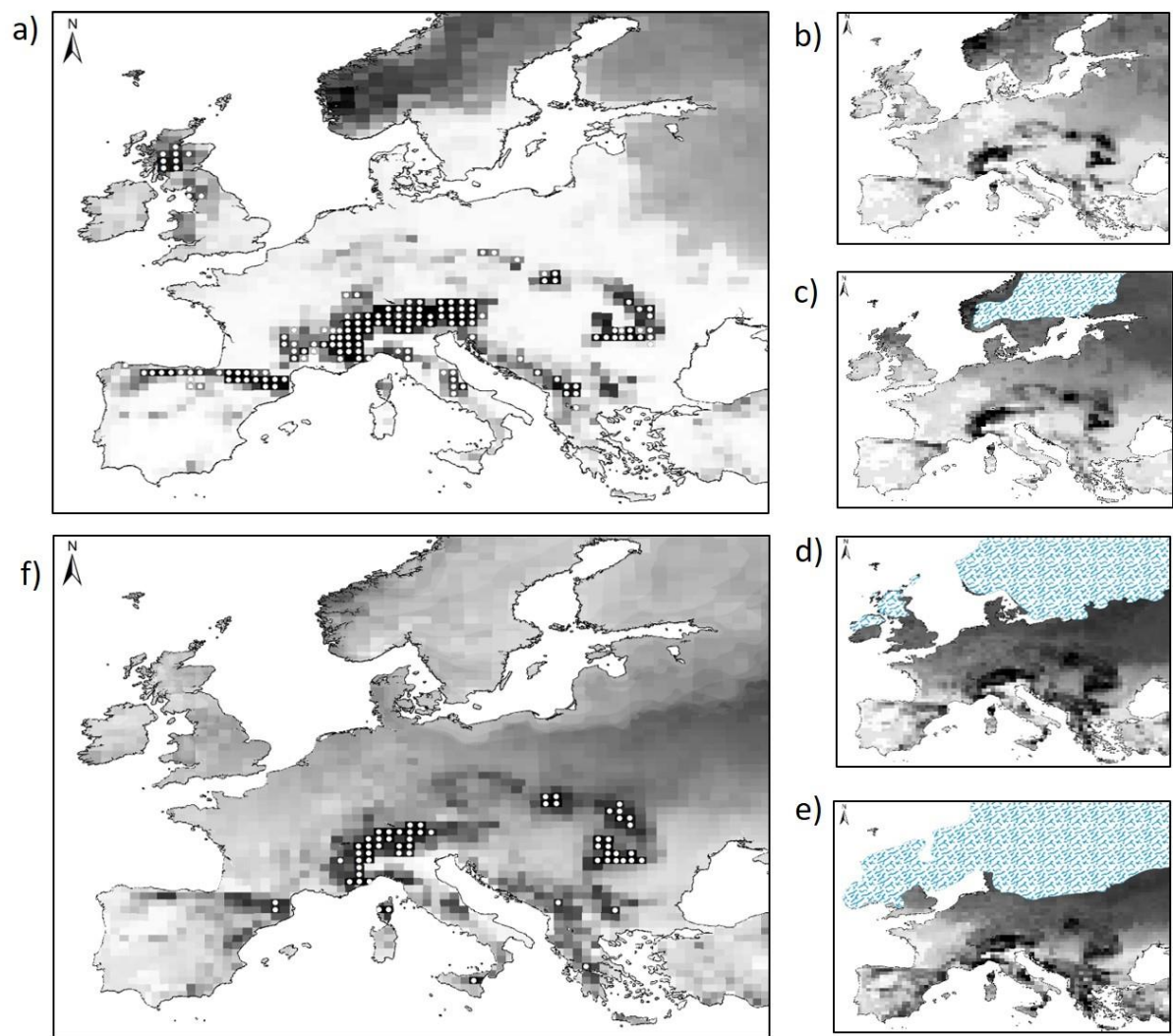


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772 **Figure 1**



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774 **Figure 2**

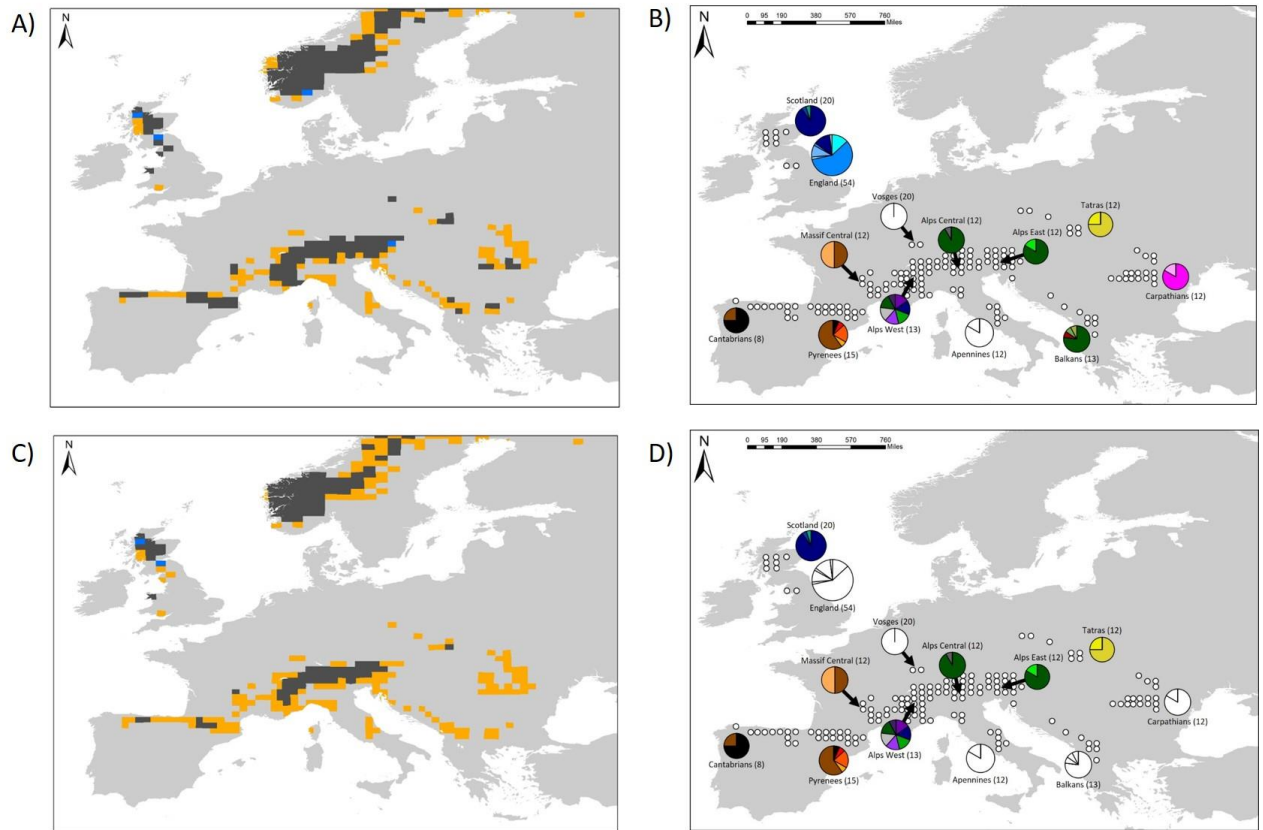


Figure 3

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Table 1

Region	Current genetic diversity				% Range change (low)	% Range change (high)	<i>Haplotypes at risk</i>	
	H_n	H_u	H_d	π			H_r (low)	H_r (high)
All	31	27	0.89	0.0055	-38.6%	-64.3%	1	12
Vosges	1	0	0	0	-100%	-100%		
Scotland	3	1	0.194	0.0003	-37.5%	-25%		
Pyrenees	5	3	0.629	0.004	-20%	-73.3%		
Massif Central	2	1	0.545	0.0008	No change	-50%		
England	7	6	0.638	0.0015	-50%	-100%		6
Carpathians	2	2	0.303	0.0005	-70.6%	-100%		2
Tatras	2	2	0.409	0.0006	-25%	-75%		
Cantabrians	2	0	0.429	0.0059	-63.6%	-81.8%		
Balkans West	4	3	0.423	0.0024	-75%	-100%		3
Apennines	2	1	0.303	0.0005	-100%	-100%	1	1
Alps West	7	5	0.912	0.0043	-14.3% (all Alps)	-41.3% (all Alps)		
Alps East	2	1	0.303	0.0005				
Alps Central	2	1	0.182	0.0006				

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813 **Figure 1:** Current distribution of genetic diversity of *E. epiphron* and historical divergence. a)
814 Frequency pie charts of haplotypes across the species' European range, including the current observed
815 distribution of *E. epiphron* (white circles; 50 km resolution) in 13 mountain regions, with number of
816 samples (individuals) in brackets. b) TCS network of all 31 identified haplotypes. Size of circle
817 represents number of individuals containing that haplotype and tick marks represent a nucleotide
818 substitution.

819 **Figure 2:** Current and past projected distributions of *E. epiphron*, a) current probability of climate
820 suitability and current distribution records (white circles). Past climate suitability b) 6,000 years ago,
821 c) 11,000 years ago, d) 16,000 years ago e) 21,000 years ago (i.e. LGM; blue shading shows the
822 extent of the ice sheet (from (Hughes et al., 2016)). Probability values of occurrence for b-e scaled
823 from 0 (unsuitable, white) to 1 (suitable, black). Panel f shows climate stability over time since the
824 LGM produced by summing 22 outputs from SDMs for the last 21,000 years, plus the output for the
825 present (summed probability values scaled from 0.73 (white) to 20 (black), with the top 30% of grids
826 shown as white circles. See Appendix S4 in Supporting Information for all output maps.

827 **Figure 3:** Projecting future climate suitability for *E. epiphron* in 2070 under two RCP climate change
828 scenarios, and associated projected loss of genetic diversity. A) low RCP 2.6 climate scenario (~1°C
829 increase by 2070), and C) high RCP 8.5 scenario (~2-3°C increase by 2070) showing grids projected
830 to remain climatically suitable (black), become unsuitable (orange), and become suitable (blue). B)
831 low RCP 2.6 scenario haplotype map with predicted lost haplotypes coloured in white (2 regions lost,
832 1 unique haplotype lost), and D) high RCP 8.5 haplotype map with predicted lost haplotypes coloured
833 in white (5 regions lost, 12 unique haplotypes lost).

834 **Table 1:** Current genetic diversity, and projected loss of climate suitability and haplotype loss in the
835 future (2070). H_n = number of haplotypes; H_u = number of unique haplotypes; π = Nei nucleotide
836 diversity (P_i); % range change = % change in range size (number of occupied 50km grid squares) in
837 the future compared with current distribution, and H_r = number of unique haplotypes at risk in the
838 future, under RCP 2.6 (low) and 8.5 (high) climate scenarios.

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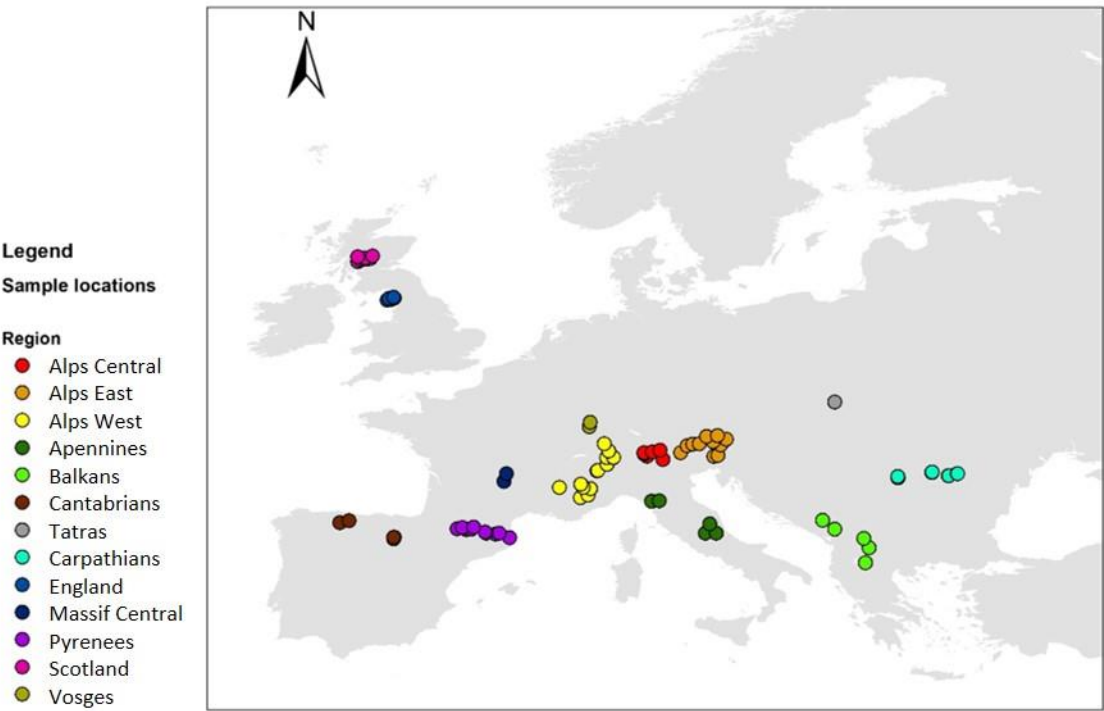
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849 **Appendix S2:** Population locations for all individuals used in mtDNA analysis from 13 mountain
850 regions.

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853 **Appendix S3:** Bioclimatic variables used in SDMs to predict climate suitability for *E. epiphron*,
854 extracted from mean temperature and precipitation data between 1970 and 2000
855 (<http://www.worldclim.org/>). 'Cells' are 2.5 arc minute (~4.5 km) resolution data extracted from
856 within a 50km grid at the same spatial extent as distribution data. 'Season' refers to mean data from
857 summer (June, July, August) and winter (December, January, February).

Climate Variable	Location in 50km grid	Season
Mean Temperature	Coldest cell	Winter (Coldest quarter)
Mean Temperature	Coldest cell	Summer (Warmest quarter)
Mean Temperature	Warmest cell	Winter (Coldest quarter)
Mean Temperature	Warmest cell	Summer (Warmest quarter)
Mean Precipitation	Wettest cell	Winter (Coldest quarter)
Mean Precipitation	Wettest cell	Summer (Warmest quarter)
Mean Precipitation	Driest cell	Winter (Coldest quarter)
Mean Precipitation	Driest cell	Summer (Warmest quarter)

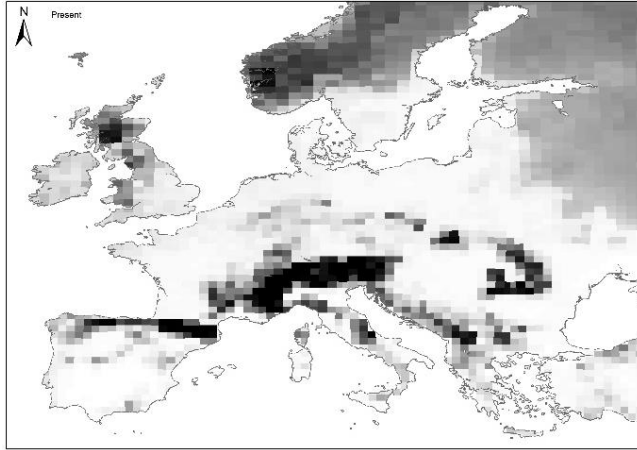
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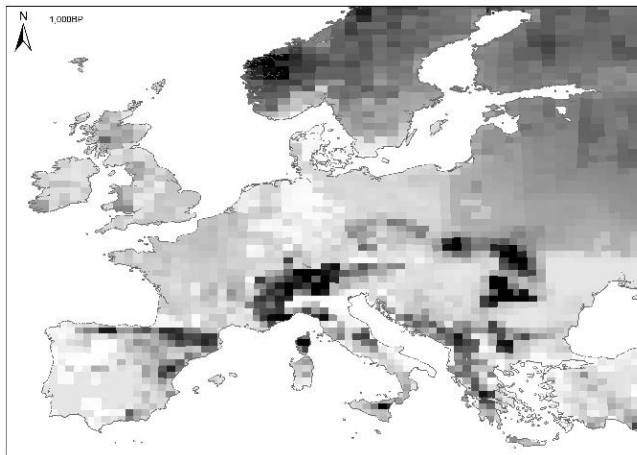
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Appendix S4: All SDM outputs showing probability of climate suitability from present-day to 21,000 years ago (22 outputs in total). Probability values of occurrence for all panels are scaled from 0 (unsuitable, white) to 1 (suitable, black). Ice sheets (from (Hughes et al., 2016), blue shading) are present from 21,000 years BP to 10,000 years BP.

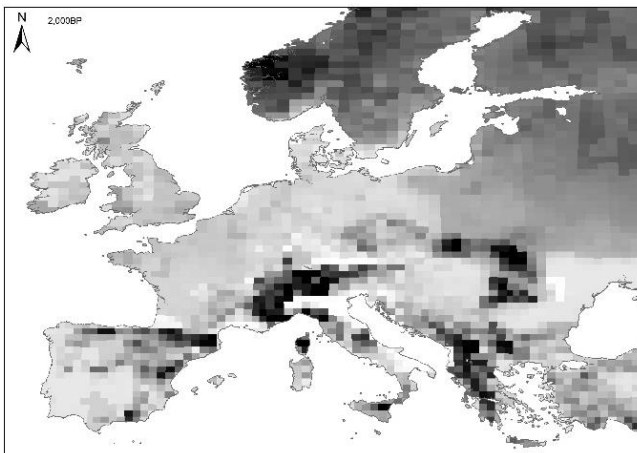
Present



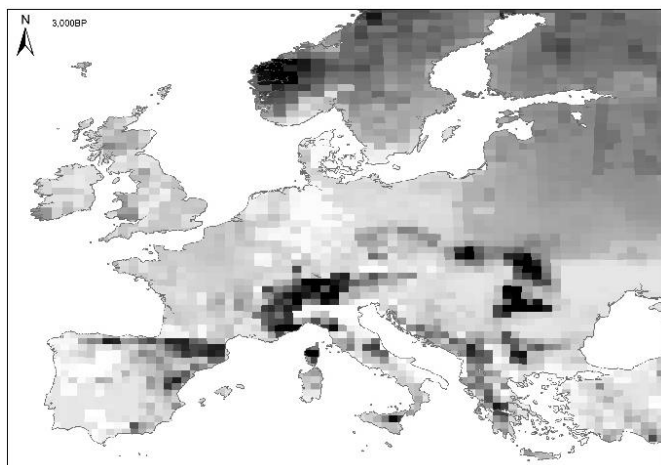
1,000 years BP



2,000 years BP

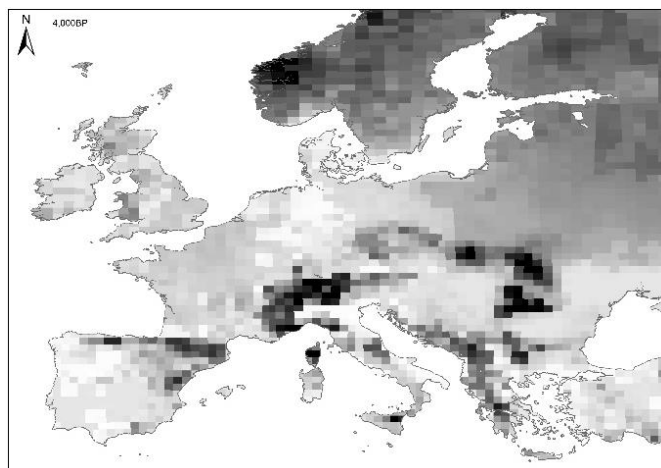


872 3,000 years BP



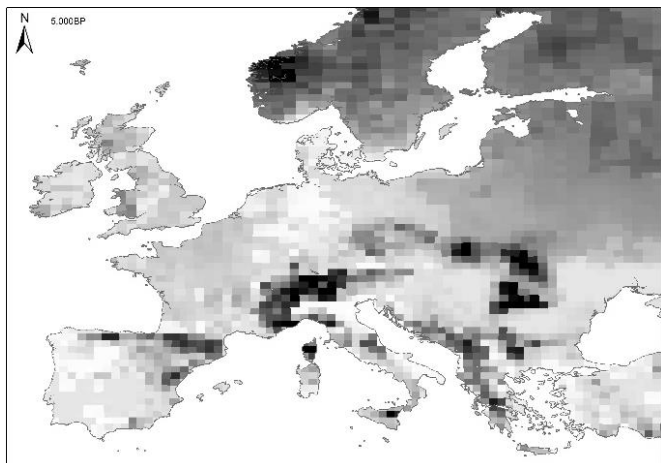
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874 4,000 years BP



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876 5,000 years BP



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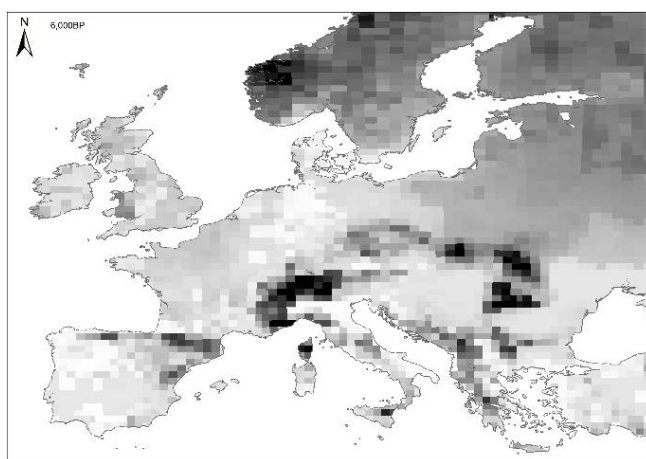
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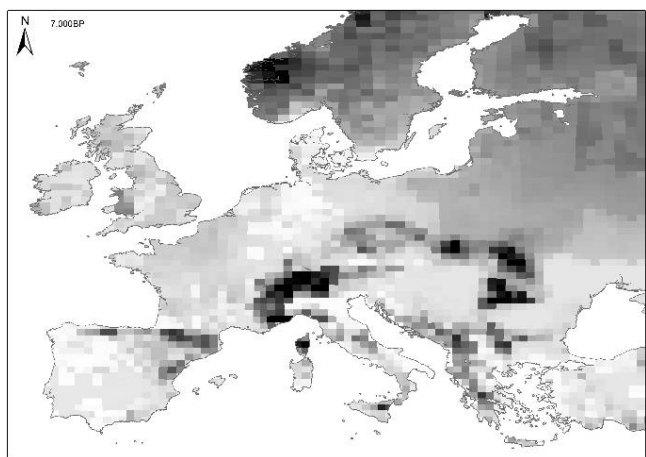
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882 6,000 years BP



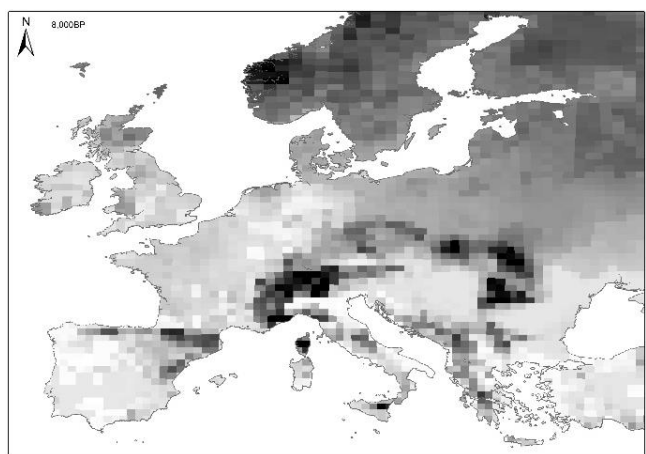
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884 7,000 years BP



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886 8,000 years BP



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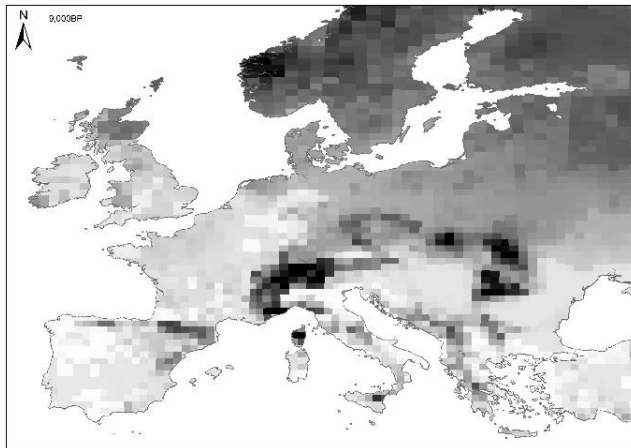
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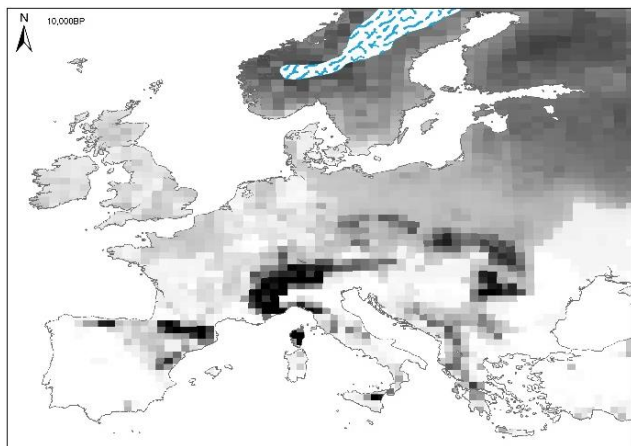
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892 9,000 years BP



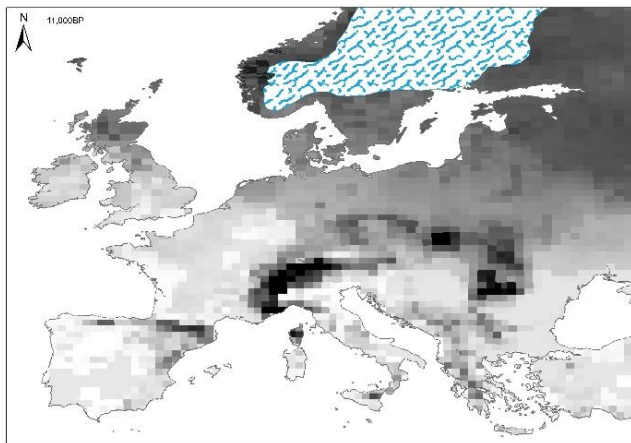
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894 10,000 years BP



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896 11,000 years BP



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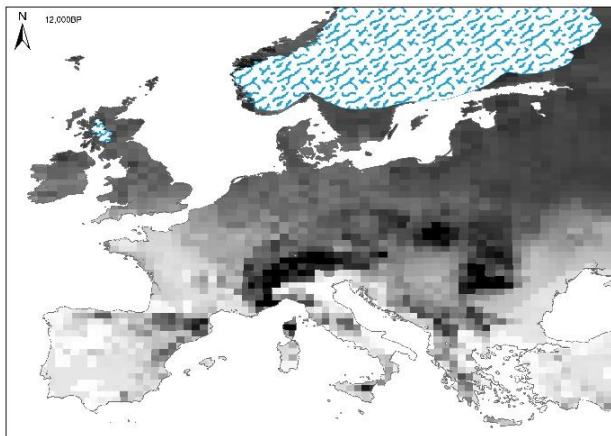
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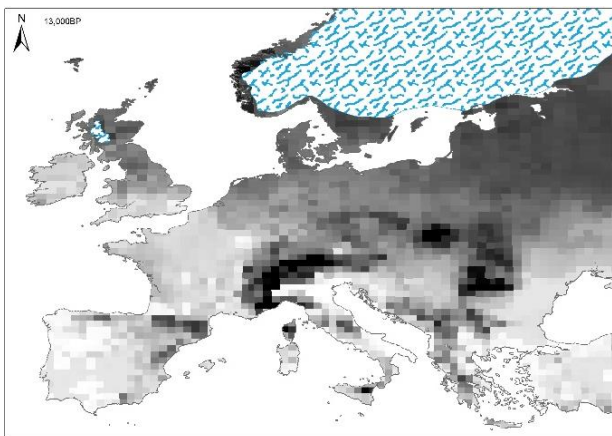
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903 12,000 years BP



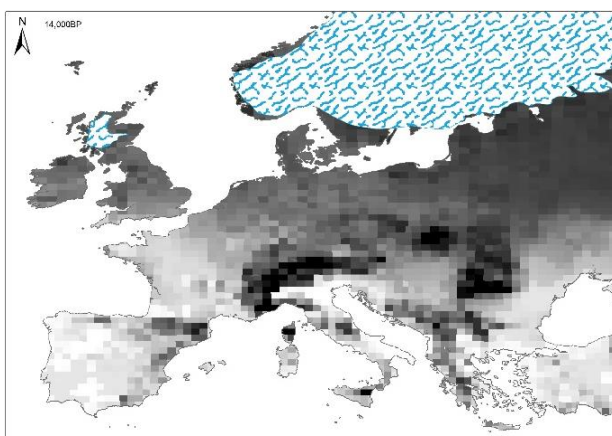
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905 13,000 years BP



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907 14,000 years BP



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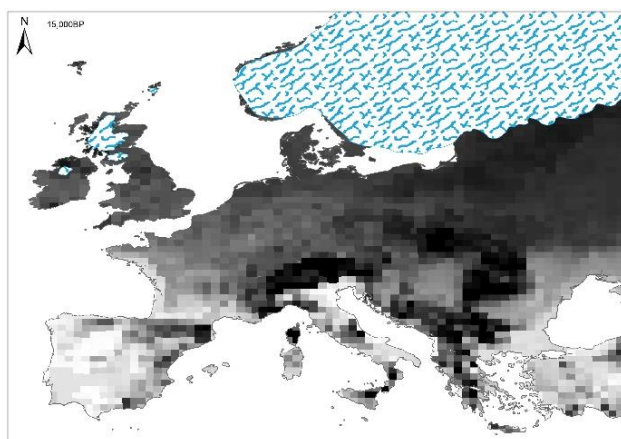
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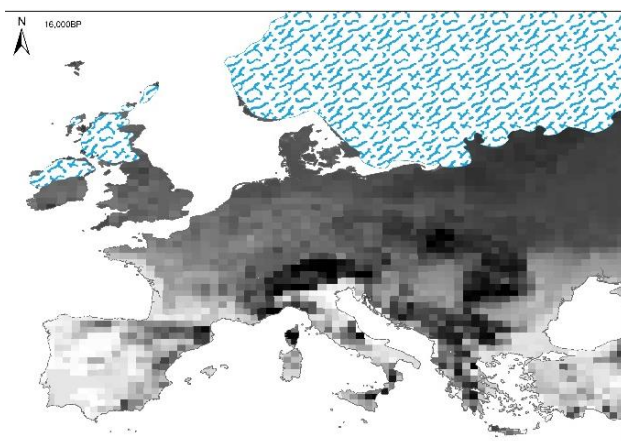
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914 15,000 years BP



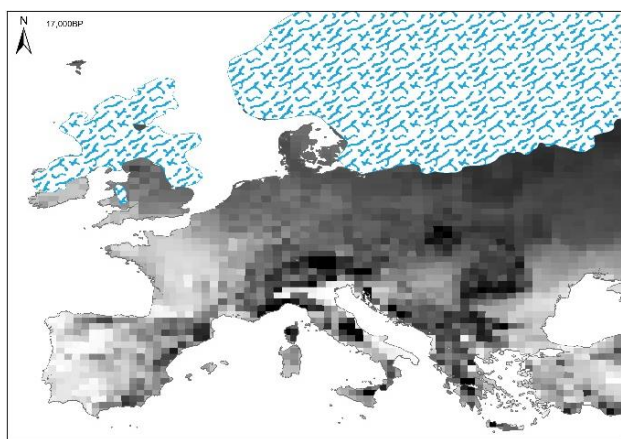
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916 16,000 years BP



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918 17,000 years BP



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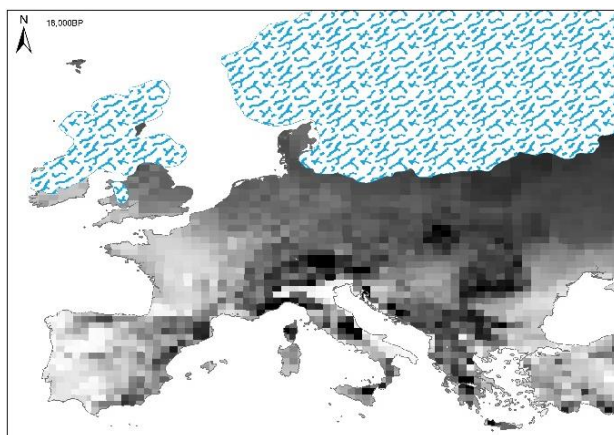
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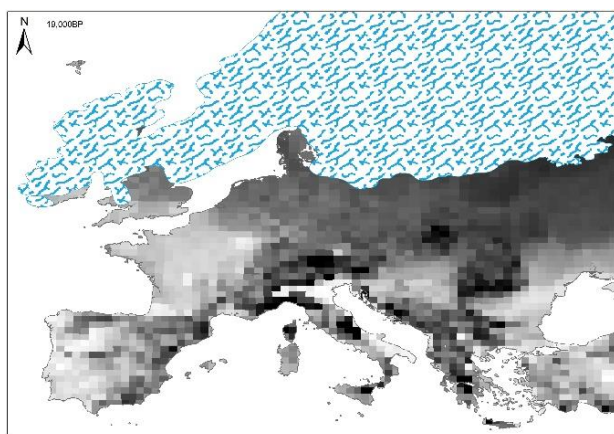
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925 18,000 years BP



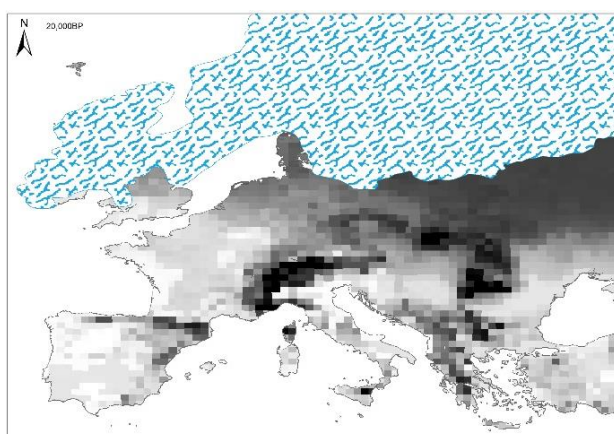
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927 19,000 years BP



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929 20,000 years BP



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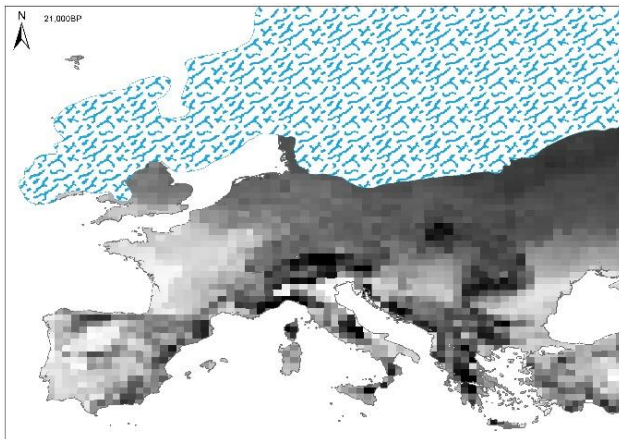
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936 21,000 years BP



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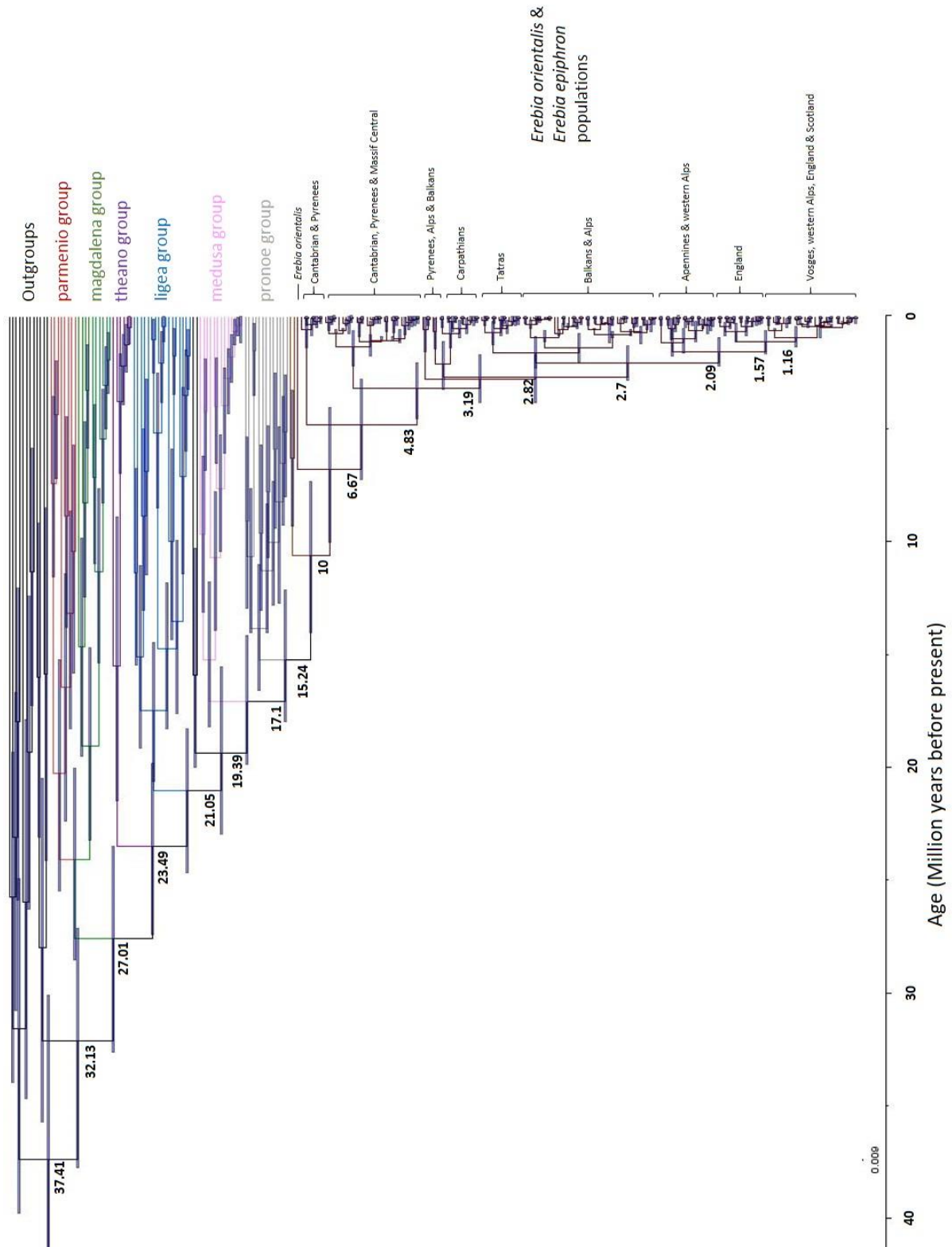
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Appendix S5: CO1 phylogenetic tree of the *Erebia* genus, outgroups and *E. epiphron* populations. Phylogenetic tree analyses were performed in Beast using methods described by Pena, Witthauer, Kleckova, Fric, & Wahlberg, (2015). Outgroup and *Erebia* genus data were accessed from Genbank using accession numbers in Pena et al., (2015). Age of split between *Erebia* and sister taxa of 37.41 Myr (Pena et al., 2015) was used to calibrate the age split between *Erebia epiphron* and *E. orientalis*. Scale bar represents age of tree in million years before present. Node number represent estimated age of node with blue error bars.

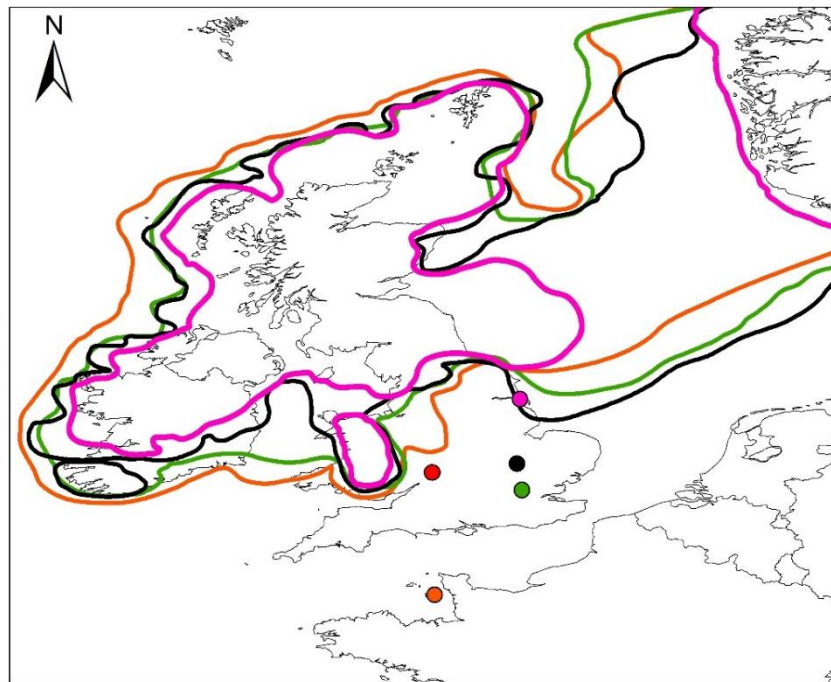


Appendix S6: Locations and ages of remains in the UK of the cool-adapted beetle species *Partobus septentrionis*, *Amara alpina*, *Amara quenseli* and *Notaris aethiops*, with corresponding most credible ice sheet extent. Beetle fossil data acquired from BugsCEP (Buckland & Buckland, 2006), ice sheet data from (Hughes, Gyllencreutz, Lohne, Mangerud, & Svendsen, 2016).

Legend

Age of beetle remains (BP)

- 18500
- 19500
- 21580
- 25500
- 26000
- Ice sheet (most credible) 18kBP
- Ice sheet (most credible) 19kBP
- Ice sheet (most credible) 21kBP
- Ice sheet (most credible) 25kBP



References

- Buckland, P. I., & Buckland, P. C. (2006). BugsCEP Coleopteran Ecology Package. *IGBP PAGES/World Data Center for Paleoclimatology Data Contribution Series*, NOAA/NCDC Paleoclimatology Program, Boulder CO, USA. <http://www.bugscep.com>.
- Hughes, A. L. C., Gyllencreutz, R., Lohne, O. S., Mangerud, J., & Svendsen, J. I. (2016). The last Eurasian ice sheets - a chronological database and time-slice reconstruction, DATED-1. *Boreas*, 45(1). doi:10.1111/bor.12142
- Pena, C., Witthauer, H., Kleckova, I., Fric, Z., & Wahlberg, N. (2015). Adaptive radiations in butterflies: evolutionary history of the genus *Erebia* (Nymphalidae: Satyrinae). *Biological Journal of the Linnean Society*, 116(2), 449-467. doi:10.1111/bij.12597

993 **Appendix S1:** Sample locations and accession numbers

994

Area	Region	Code	Locality	LATITUDE	LONGITUDE	Haplotype	Genbank Accession No.	BOLD Process ID
Europe	Alps Central	10-1_AlpsC_4	Sellajoch, Italy	46.50	9.87	1		
UK	Scotland	10-1_Scot_6	Ben Lawers, Perth and Kinross, UK	56.53	-4.25	8		
Europe	Alps Central	10-2_AlpsC_5	Sellajoch, Italy	46.50	9.87	1		
Europe	Alps Central	10-3_AlpsC_8	Sellajoch, Italy	46.50	9.87	1		
UK	Lake District	1-1_Lakes_1	Irton fell, Cumbria, UK	54.41	-3.32	4		
UK	Scotland	11-1_Scot_7	Beinn Odhar, Tyndrum, UK	56.46	-4.69	8		
Europe	Alps East	11-4_AlpsE_1	Rein in Taufers, Italy	46.95	12.07	1		
UK	Lake District	1-2_Lakes_2	Irton fell, Cumbria, UK	54.41	-3.32	4		
Europe	Alps East	12-1_AlpsE_3	Hochköng, Austria	47.42	13.05	1		
UK	Scotland	12-1_Scot_8	Ben Lawers, Perth and Kinross, UK	56.56	-4.17	8		
Europe	Apennines	13-1_Apen_5	Prati di Tivo, Italy	42.47	13.55	10		
UK	Scotland	13-1_Scot_9	Schiehallion, Perth and Kinross, UK	56.67	-4.07	8		
Europe	Apennines	13-2_Apen_6	Prati di Tivo, Italy	42.47	13.55	10		
Europe	Apennines	13-3_Apen_7	Prati di Tivo, Italy	42.47	13.55	10		
Europe	Apennines	13-4_Apen_8	Prati di Tivo, Italy	42.47	13.55	10		
Europe	Apennines	14-1_Apen_1	Terminillo, Italy	42.47	13.00	10		
Europe	Apennines	14-2_Apen_2	Terminillo, Italy	42.47	13.00	10		
Europe	Apennines	14-3_Apen_3	Terminillo, Italy	42.47	13.00	10		
Europe	Apennines	14-4_Apen_4	Terminillo, Italy	42.47	13.00	10		
Europe	Pyrenees	15-1_Pyr_2	Candanchu, Spain	42.75	0.53	14		
Europe	Alps East	16-1_AlpsE_4	Sölkpass, Austria	47.27	14.07	1		
Europe	Alps Central	17-1_AlpsC_6	Thanai, Italy	46.72	10.67	1		
Europe	Alps Central	17-2_AlpsC_7	Thanai, Italy	46.72	10.67	1		
Europe	Alps East	18-1_AlpsE_5	Schönfeld, Austria	46.98	13.78	1		
Europe	Alps East	19-1_AlpsE_2	Sajatmähder, Aiustria	47.03	12.35	1		
Europe	Alps Central	2-1_AlpsC_1	Berninapass, Switzerland	46.40	10.02	1		
UK	Lake District	2-1_Lakes_3	Grisedale, Cumbria, UK	54.52	-2.95	3		
Europe	Carpathians	21-1_CarpS_2	Valea Caprei, Romania	45.58	24.62	19		
Europe	Alps Central	2-2_AlpsC_2	Berninapass, Switzerland	46.40	10.02	1		
UK	Lake District	2-2_Lakes_4	Grisedale, Cumbria, UK	54.52	-2.95	3		
Europe	Alps West	22- 1_AlpsW_3	Passo del Monte Moro, Italy	45.98	7.97	1		
Europe	Alps Central	2-3_AlpsC_3	Berninapass, Switzerland	46.40	10.02	1		
Europe	Pyrenees	23-1_Pyr_1	Panticosa, Spain	42.68	0.27	13		
Europe	Pyrenees	24-1_Pyr_4	Canigou, France	42.47	2.42	16		
Europe	Vosges	26-1_Vosg_7	Markstein, France	47.92	7.04	8		

Europe	Vosges	26-2_Vosg_8	Markstein, France	47.92	7.04	8		
Europe	Vosges	26-3_Vosg_9	Markstein, France	47.92	7.04	8		
Europe	Vosges	26-4_Vosg_10	Markstein, France	47.92	7.04	8		
Europe	Vosges	26-5_Vosg_11	Markstein, France	47.92	7.04	8		
Europe	Vosges	26-6_Vosg_12	Markstein, France	47.92	7.04	8		
Europe	Vosges	27-1_Vosg_1	Col du Calvaire, France	48.14	7.10	8		
Europe	Vosges	27-2_Vosg_2	Col du Calvaire, France	48.14	7.10	8		
Europe	Vosges	27-3_Vosg_3	Col du Calvaire, France	48.14	7.10	8		
Europe	Vosges	27-4_Vosg_4	Col du Calvaire, France	48.14	7.10	8		
Europe	Vosges	27-5_Vosg_5	Col du Calvaire, France	48.14	7.10	8		
Europe	Vosges	27-6_Vosg_6	Col du Calvaire, France	48.14	7.10	8		
Europe	Tatras	28-1_CarpN_1	Babky, Tatra Mts, Slovakia	49.18	19.63	17		
Europe	Tatras	28-10_CarpN_10	Babky, Tatra Mts, Slovakia	49.18	19.63	17		
Europe	Tatras	28-11_CarpN_11	Babky, Tatra Mts, Slovakia	49.18	19.63	17		
Europe	Tatras	28-12_CarpN_12	Babky, Tatra Mts, Slovakia	49.18	19.63	18		
Europe	Tatras	28-2_CarpN_2	Babky, Tatra Mts, Slovakia	49.18	19.63	17		
Europe	Tatras	28-3_CarpN_3	Babky, Tatra Mts, Slovakia	49.18	19.63	17		
Europe	Tatras	28-4_CarpN_4	Babky, Tatra Mts, Slovakia	49.18	19.63	17		
Europe	Tatras	28-5_CarpN_5	Babky, Tatra Mts, Slovakia	49.18	19.63	17		
Europe	Tatras	28-6_CarpN_6	Babky, Tatra Mts, Slovakia	49.18	19.63	18		
Europe	Tatras	28-7_CarpN_7	Babky, Tatra Mts, Slovakia	49.18	19.63	17		
Europe	Tatras	28-8_CarpN_8	Babky, Tatra Mts, Slovakia	49.18	19.63	18		
Europe	Tatras	28-9_CarpN_9	Babky, Tatra Mts, Slovakia	49.18	19.63	17		
Europe	Carpathians	29-4_CarpS_1	Retezat, Romania	45.37	22.87	19		
Europe	Balkans	30-1_Balk_1	Kom vasjeviak, Montenegro	42.68	19.63	1		
Europe	Balkans	30-2_Balk_2	Kom vasjeviak, Montenegro	42.68	19.63	1		
Europe	Balkans	30-3_Balk_3	Kom vasjeviak, Montenegro	42.68	19.63	1		
Europe	Balkans	30-4_Balk_4	Kom vasjeviak, Montenegro	42.68	19.63	1		
Europe	Alps West	3-1_AlpsW_2	Täschalp, Switzerland	47.05	7.82	1		
UK	Lake District	3-1_Lakes_5	Raise, Cumbria, UK	54.55	-3.00	7		
Europe	Balkans	31-1_Balk_5	Vjetrena brda, Durmitar, Montenegro	43.12	19.02	1		
Europe	Balkans	31-2_Balk_6	Vjetrena brda, Durmitar, Montenegro	43.12	19.02	1		
Europe	Balkans	31-3_Balk_7	Vjetrena brda, Durmitar, Montenegro	43.12	19.02	1		
Europe	Balkans	31-4_Balk_8	Vjetrena brda, Durmitar, Montenegro	43.12	19.02	1		
Europe	Balkans	31-5_Balk_9	Vjetrena brda, Durmitar, Montenegro	43.12	19.02	1		
UK	Lake District	3-2_Lakes_6	Raise, Cumbria, UK	54.55	-3.00	3		
UK	Lake District	4-1_Lakes_7	Langdale, Cumbria, UK	54.46	-3.10	4		
Europe	Pyrenees	4-1_Pyr_3	Étang d'Areau, France	42.77	1.12	15		
UK	Lake District	4-2_Lakes_8	Langdale, Cumbria, UK	54.46	-3.10	5		
Europe	Alps West	5-1_AlpsW_1	Grindelwald, Switzerland	46.67	8.03	8		

UK	Lake District	5-1_Lakes_9	Wynrose, Cumbria, UK	54.42	-3.13	6		
UK	Lake District	5-2_Lakes_10	Wynrose, Cumbria, UK	54.42	-3.13	6		
Europe	Massif Central	6-1_MasC_7	Puy Mary, France	45.52	2.80	16		
UK	Scotland	6-1_Scot_1	Glen Lyon, Perth and Kinross, UK	56.58	-4.44	8		
Europe	Massif Central	6-2_MasC_8	Puy Mary, France	45.52	2.80	16		
Europe	Massif Central	6-3_MasC_9	Puy Mary, France	45.52	2.80	16		
Europe	Massif Central	6-4_MasC_10	Puy Mary, France	45.52	2.80	16		
Europe	Massif Central	6-5_MasC_11	Puy Mary, France	45.52	2.80	16		
Europe	Massif Central	7-1_MasC_1	Puy de Sancy/Chastraix-sancy, France	45.52	2.80	29		
UK	Scotland	7-1_Scot_2	Ben Lui, Tyndrum, UK	56.39	-4.83	8		
Europe	Massif Central	7-2_MasC_2	Puy de Sancy/Chastraix-sancy, France	45.52	2.80	29		
Europe	Massif Central	7-3_MasC_3	Puy de Sancy/Chastraix-sancy, France	45.52	2.80	29		
Europe	Massif Central	7-4_MasC_4	Puy de Sancy/Chastraix-sancy, France	45.52	2.80	29		
Europe	Massif Central	7-5_MasC_5	Puy de Sancy/Chastraix-sancy, France	45.52	2.80	29		
Europe	Massif Central	7-6_MasC_6	Puy de Sancy/Chastraix-sancy, France	45.52	2.80	29		
UK	Scotland	8-1_Scot_3	Ben Lui, Tyndrum, UK	56.38	-4.81	8		
Europe	Alps East	9-1_AlpsE_6	Mangart, Slovenia	46.45	13.65	9		
UK	Scotland	9-1_Scot_4	Glencoe, Argyll, UK	56.63	-4.85	8		
Europe	Alps East	9-2_AlpsE_7	Mangart, Slovenia	46.45	13.65	9		
UK	Scotland	9-2_Scot_5	Glencoe, Argyll, UK	56.63	-4.85	8		
Europe	Carpathians	RVcoll06M974	Săcele, Braşov, Romania	45.52	25.92	19	HQ004371	EZROM149-08
Europe	Carpathians	RVcoll06M985	Măneciu, Prahova, Romania	45.52	25.93	19	HQ004369	EZROM672-08
Europe	Carpathians	RVcoll06M987	Măneciu, Prahova, Romania	45.52	25.93	19	HQ004373	EZROM914-08
Europe	Carpathians	RVcoll06V683	Râu de Mori, Hunedoara, Romania	45.30	22.87	19	HQ004372	EZROM150-08
Europe	Carpathians	RVcoll06V706	Uricani, Hunedoara, Romania	45.31	22.88	19	GU669667	EZROM1037-09
Europe	Carpathians	RVcoll07D631	Buşteni, Prahova, Romania	45.40	25.48	20	HQ004374	EZROM915-08
Europe	Carpathians	RVcoll07E456	Moroeni, Dâmboviţa, Romania	45.40	25.47	20	HQ004370	EZROM151-08
Europe	Carpathians	RVcoll07E495	Uricani, Hunedoara, Romania	45.30	22.88	19	HQ004375	EZROM916-08
Europe	Pyrenees	RVcoll07W121	Vielha e Mijaran, Lleida, Spain	42.66	0.75	14	GU669854	EZSPC381-09
Europe	Carpathians	RVcoll08M607	Arefu, Argeş, Romania	45.59	24.63	19	HQ004377	EZROM917-08
Europe	Carpathians	RVcoll08M614	Arefu, Argeş, Romania	45.59	24.63	19	HQ004376	EZROM918-08
Europe	Pyrenees	RVcoll08M994	El Pas de la Casa, Encamp, Andorra	42.54	1.70	16	HM901314	EZSPC1113-10
Europe	Pyrenees	RVcoll08M995	El Pas de la Casa, Encamp, Andorra	42.54	1.70	16	HM901315	EZSPC1114-10
Europe	Pyrenees	RVcoll08R174	Setcases, Girona, Spain	42.43	2.24	16	GU669853	EZSPC380-09
Europe	Pyrenees	RVcoll08R257	Alt Àneu, Lleida, Spain	42.67	0.99	16	HM901357	EZSPC1157-10
Europe	Pyrenees	RVcoll08R259	Alt Àneu, Lleida, Spain	42.67	0.99	16	GU669855	EZSPC382-09

Europe	Pyrenees	RVcoll08R260	Alt Àneu, Lleida, Spain	42.67	0.99	16	GU669848	EZSPC383-09
Europe	Pyrenees	RVcoll08R261	Alt Àneu, Lleida, Spain	42.67	0.99	16	GU669849	EZSPC384-09
Europe	Cantabrian	RVcoll08R410	Lena, Asturias, Spain	43.00	-5.76	16	GU675818	EZSPM221-09
Europe	Cantabrian	RVcoll08R413	Caso, Asturias, Spain	43.11	-5.27	16	GU675815	EZSPM223-09
Europe	Pyrenees	RVcoll09T080	Vielha e Mijaran, Lleida	42.67	0.73	14	JF847985	EZSPN036-09
Europe	Pyrenees	RVcoll09X029	Meranges, Girona	42.47	1.76	16	HM901499	EZSPC1365-10
Europe	Alps West	RVcoll10B939	Uvernet-Fours, Alpes-de-Haute-Provence	44.29	6.59	23	DQ338778	WMB2684-13
Europe	Alps West	RVcoll10C021	Arvieux, Hautes-Alpes, France	44.82	6.74	25	KR138782	WMB2693-13
Europe	Alps West	RVcoll11I916	Chichilianne, Isère, France	44.81	5.52	8	KP870625	EULEP170-14
Europe	Alps Central	RVcoll11J460	Bever, Grisons, Switzerland	46.55	9.85	1	KP870445	EULEP183-14
UK	Lake District	RVcoll12R462	Cockermouth, Cumbria, UK	54.50	-3.21	4	KP870916	EULEP261-14
UK	Lake District	RVcoll12R463	Cockermouth, Cumbria, UK	54.50	-3.21	4	KP870577	EULEP261-14
UK	Lake District	RVcoll12R464	Cockermouth, Cumbria, UK	54.50	-3.21	4	KP870587	EULEP262-14
UK	Lake District	RVcoll12R465	Cockermouth, Cumbria, UK	54.50	-3.21	4	KP870931	EULEP263-14
UK	Lake District	RVcoll12R466	Cockermouth, Cumbria, UK	54.50	-3.21	4	MK155216	EULEP264-14
UK	Scotland	RVcoll12R468	Killin, Stirling, UK	56.51	-4.50	8	KP870980	EULEP265-14
UK	Scotland	RVcoll12R469	Killin, Stirling, UK	56.51	-4.50	8	KP870580	EULEP266-14
UK	Scotland	RVcoll12R471	Killin, Stirling, UK	56.51	-4.50	8	KP870616	EULEP267-14
Europe	Apennines	RVcoll14A259	Ussita, Macerata, Italy	42.94	13.22	10	MK155192	EULEP1875-15
Europe	Apennines	RVcoll14A260	Ussita, Macerata, Italy	42.94	13.22	10	KR138751	WMB5256-14
Europe	Apennines	RVcoll14A446	Abetone, Pistoia, Italy	44.13	10.64	11	MK155190	EULEP1878-15
Europe	Apennines	RVcoll14A619	Massa, Lucca, Italy	44.10	10.23	11	KR138798	WMB5276-14
Europe	Alps West	RVcoll14D994	Villar Pellice, Turin, Italy	44.75	7.11	22	MK155199	BIBSA206-15
Europe	Alps West	RVcoll14E100	Acceglio, Cuneo, Italy	44.43	6.98	23	MK155180	BIBSA298-15
Europe	Alps West	RVcoll14I053	Saint-Marcel, Aosta, Italy	45.66	7.44	21	MK155204	BIBSA385-15
Europe	Alps West	RVcoll14I060	Saint-Marcel, Aosta, Italy	45.69	7.48	21	MK155214	BIBSA392-15
Europe	Massif Central	RVcoll14J771	Le Falgoux, Cantal, France	45.11	2.66	16	MK155198	EULEP2115-15
Europe	Alps West	RVcoll14N049	Parco Veglia Devero, Italy	46.34	8.28	24		
Europe	Cantabrian	RVcoll14N230	Villavelayo, La Rioja, Spain	42.18	-3.00	12	HE614683	WMB5455-14
Europe	Balkans	RVcoll14N877	Ljuboten	42.20	21.13	27	MK155194	EULEP2793-15
Europe	Alps Central	RVcoll14O005	Pradalago, Trentino, Italy	46.25	10.81	2		
Europe	Cantabrian	RVcoll15D777	Ezcaray, La Rioja, Spain	42.26	-2.98	12	MK155181	EULEP5633-17
Europe	Cantabrian	RVcoll15D778	Ezcaray, La Rioja, Spain	42.26	-2.98	12	MK155185	EULEP5634-17
Europe	Cantabrian	RVcoll15D779	Pazuengos, La Rioja, Spain	42.25	-2.95	12	MK155187	EULEP5635-17
Europe	Cantabrian	RVcoll15D780	San Millán de Cogolla, La Rioja, Spain	42.25	-2.94	12	MK155210	EULEP5636-17
Europe	Cantabrian	RVcoll15D781	Pazuengos, La Rioja, Spain	42.25	-2.95	12	MK155217	EULEP5637-17

Europe	Pyrenees	RVcoll15D782	Fresneda de la Sierra Tirón, Burgos, Spain	42.24	2.97	12	MK155191	EULEP5638-17
Europe	Alps West	RVcoll15H312	Mund, Valais, Switzerland	46.33	7.94	24	MK155202	EULEP5649-17
Europe	Alps East	RVcoll15I016	Chiusaforte, Udine, Italy	46.41	13.44	1	MK155179	EULEP5650-17
Europe	Alps East	RVcoll15I330	Kals am Großglockner, Tyrol, Austria	47.04	12.69	1	MK155215	EULEP5651-17
Europe	Alps East	RVcoll15I602	Ramsau am Dachstein, Styria, Austria	47.46	13.62	1	MK155211	EULEP5652-17
Europe	Alps East	RVcoll15I860	Muhr, Salzburg, Austria	47.15	13.38	1	MK155178	EULEP3794-16
Europe	Alps Central	RVcoll15I957	La Punt-Chamues-ch, Grisons, Switzerland	46.58	9.84	1	MK155212	EULEP3795-16
Europe	Alps Central	RVcoll15J040	Tschier, Grisons, Switzerland	46.63	10.29	1	MK155213	EULEP3796-16
Europe	Alps West	RVcoll15J516	Val-des-Prés, Hautes-Alpes	44.97	6.61	22	MK155197	EULEP3797-16
Europe	Alps East	RVcoll15K528	Santa Cristina Gherdëina, Bolzano, Italy	46.60	11.74	1	MK155205	BIBSA1077-15
Europe	Balkans	RVcoll15P093	Studeničani, Skopje	41.73	21.40	28	MK155193	EULEP3798-16
Europe	Balkans	RVcoll15P094	Pelister Mt. (Gol. Ez. - Or. Bar.)	40.96	21.20	1		
Europe	Balkans	RVcoll15Q015	Shar Mts. (prema vrv Ljuboten)	42.20	21.13	26		
UK	Lake District	01_EE_snps	Kidsty Pike/High Raise, Cumbria, UK	54.51	-2.87	4		
UK	Lake District	02_EE_snps	Kidsty Pike/High Raise, Cumbria, UK	54.51	-2.87	8		
UK	Lake District	03_EE_snps	Kidsty Pike/High Raise, Cumbria, UK	54.51	-2.87	8		
UK	Lake District	04_EE_snps	Kidsty Pike/High Raise, Cumbria, UK	54.51	-2.87	4		
UK	Lake District	14_EE_snps	Kidsty Pike/High Raise, Cumbria, UK	54.51	-2.87	4		
UK	Lake District	11_EE_snps	Kidsty Pike/High Raise, Cumbria, UK	54.51	-2.87	4		
UK	Lake District	13_EE_snps	Kidsty Pike/High Raise, Cumbria, UK	54.51	-2.87	4		
UK	Lake District	15_EE_snps	Kidsty Pike/High Raise, Cumbria, UK	54.51	-2.87	4		
UK	Lake District	17_EE_snps	Kidsty Pike/High Raise, Cumbria, UK	54.51	-2.87	4		
UK	Lake District	16_EE_snps	Kidsty Pike/High Raise, Cumbria, UK	54.51	-2.87	4		
UK	Lake District	18_EE_snps	Kidsty Pike/High Raise, Cumbria, UK	54.51	-2.87	4		
UK	Lake District	21_EE_snps	Irton fell, Cumbria, UK	54.41	-3.33	4		
UK	Lake District	22_EE_snps	Irton fell, Cumbria, UK	54.41	-3.33	4		
UK	Lake District	12_EE_snps	Kidsty Pike/High Raise, Cumbria, UK	54.51	-2.87	4		
UK	Lake District	24_EE_snps	Irton fell, Cumbria, UK	54.41	-3.33	4		
UK	Lake District	26_EE_snps	Irton fell, Cumbria, UK	54.41	-3.33	4		
UK	Lake District	25_EE_snps	Irton fell, Cumbria, UK	54.41	-3.33	4		
UK	Lake District	29_EE_snps	Irton fell, Cumbria, UK	54.41	-3.33	4		
UK	Lake District	28_EE_snps	Irton fell, Cumbria, UK	54.41	-3.33	4		
UK	Lake District	27_EE_snps	Irton fell, Cumbria, UK	54.41	-3.33	4		
UK	Lake District	23_EE_snps	Irton fell, Cumbria, UK	54.41	-3.33	4		

UK	Lake District	30_EE_snps	Irton fell, Cumbria, UK	54.41	-3.33	4		
UK	Lake District	38_EE_snps	Fleetwith (Wasdale Scree), Cumbria, UK	54.51	-3.22	4		
UK	Lake District	36_EE_snps	Fleetwith (Wasdale Scree), Cumbria, UK	54.51	-3.22	4		
UK	Lake District	39_EE_snps	Fleetwith (Wasdale Scree), Cumbria, UK	54.51	-3.22	4		
UK	Lake District	35_EE_snps	Fleetwith (Wasdale Scree), Cumbria, UK	54.51	-3.22	6		
UK	Lake District	40_EE_snps	Fleetwith (Wasdale Scree), Cumbria, UK	54.51	-3.22	6		
UK	Lake District	37_EE_snps	Fleetwith (Wasdale Scree), Cumbria, UK	54.51	-3.22	6		
UK	Lake District	05_EE_snps	Kidsty Pike/High Raise, Cumbria, UK	54.51	-2.87	8		
UK	Lake District	06_EE_snps	Kidsty Pike/High Raise, Cumbria, UK	54.51	-2.87	8		
UK	Lake District	08_EE_snps	Kidsty Pike/High Raise, Cumbria, UK	54.51	-2.87	8		
UK	Lake District	09_EE_snps	Kidsty Pike/High Raise, Cumbria, UK	54.51	-2.87	8		
UK	Lake District	20_EE_snps	Kidsty Pike/High Raise, Cumbria, UK	54.51	-2.87	8		
UK	Lake District	10_EE_snps	Kidsty Pike/High Raise, Cumbria, UK	54.51	-2.87	31		
UK	Lake District	19_EE_snps	Kidsty Pike/High Raise, Cumbria, UK	54.51	-2.87	31		
UK	Lake District	31_EE_snps	Kirkstone Pass (Troutbeck), Cumbria, UK	54.47	-2.91	3		
UK	Lake District	32_EE_snps	Kirkstone Pass (Troutbeck), Cumbria, UK	54.47	-2.91	3		
UK	Lake District	33_EE_snps	Kirkstone Pass (Troutbeck), Cumbria, UK	54.47	-2.91	3		
UK	Lake District	34_EE_snps	Kirkstone Pass (Troutbeck), Cumbria, UK	54.47	-2.91	3		
UK	Scotland	Scot_6-2	Glen Lyon, Perth and Kinross, UK	56.58	-4.44	10		
UK	Scotland	Scot_6-3	Glen Lyon, Perth and Kinross, UK	56.58	-4.44	30		
UK	Scotland	Scot_7-2	Ben Lui, Tyndrum, UK	56.39	-4.83	8		
UK	Scotland	Scot_7-3	Ben Lui, Tyndrum, UK	56.39	-4.83	8		
UK	Scotland	Scot_18-1	Beinn Chaorach, Stirling, UK	56.45	-4.68	8		
UK	Scotland	Scot_18-2	Beinn Chaorach, Stirling, UK	56.45	-4.68	8		
UK	Scotland	Scot_19-1	Stob Mhic Mhartaigh, Kinlochleven, UK	56.67	-4.94	8		
UK	Scotland	Scot_19-2	Stob Mhic Mhartaigh, Kinlochleven, UK	56.67	-4.94	8		
Europe	Vosges	Vosg_26-7	Markstein, France	47.92	7.04	8		
Europe	Vosges	Vosg_26-8	Markstein, France	47.92	7.04	8		
Europe	Vosges	Vosg_26-9	Markstein, France	47.92	7.04	8		
Europe	Vosges	Vosg_26-10	Markstein, France	47.92	7.04	8		
Europe	Vosges	Vosg_27-7	Col du Calvaire, France	48.14	7.10	8		
Europe	Vosges	Vosg_27-8	Col du Calvaire, France	48.14	7.10	8		
Europe	Vosges	Vosg_27-9	Col du Calvaire, France	48.14	7.10	8		
Europe	Vosges	Vosg_27-10	Col du Calvaire, France	48.14	7.10	8		

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